



Citation: M. Khajavi, M. Rahaie, A.Ebrahimi (2019) The effect of TiO₂ and SiO₂ nanoparticles and salinity stress on expression of genes involved in parthenolide biosynthesis in Feverfew (*Tanacetum parthenium* L.). *Caryologia* 72(1): 3-14. doi: 10.13128/caryologia-246

Received: 30th July 2018

Accepted: 24th December 2018

Published: 10th May 2019

Copyright: © 2019 M. Khajavi, M. Rahaie, A. Ebrahimi. This is an open access, peer-reviewed article published by Firenze University Press (<http://www.fupress.com/caryologia>) and distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Competing Interests: The Author(s) declare(s) no conflict of interest.

The effect of TiO₂ and SiO₂ nanoparticles and salinity stress on expression of genes involved in parthenolide biosynthesis in Feverfew (*Tanacetum parthenium* L.)

MAHSHID KHAJAVI¹, MAHDI RAHAIE^{2,*}, ASA EBRAHIMI¹

¹ Department of Biotechnology and Plant Breeding, Faculty of Agriculture, Science and Research Branch of Islamic Azad university, Tehran, Iran

² Department of Life Science Engineering, Faculty of New Sciences and Technologies, University of Tehran, Tehran, Iran

* Corresponding author: Tel: +98-21-86093408; Fax: +98-21-88497324; Email: mrahaie@ut.ac.ir

Abstract. Medicinal plants can produce various chemical compounds as secondary metabolites that have benefit to human. Feverfew (*Tanacetum parthenium* L.) is a medicinal plant belongs to the Asteraceae family. This plant due to have parthenolide compounds has attracted much attention for medicinal value and pharmacological activity. Due to the economic importance of the plant metabolite in cancer and migraine treatment, application of approaches for increasing the metabolite was the objective of this study. For this purpose, after cultivation in greenhouse, plants were treated with TiO₂ and SiO₂ nanoparticles and salinity stress at different times and concentrations. Real Time PCR used to evaluate the expression of *TpGAS*, *COST* and *TpCarS* genes which involved in secondary metabolites biosynthesis pathway (parthenolide and β-caryophyllen). It was found, SiO₂ NPs can increase the expression of *TpCarS*, *COST* and *TpGAS* in the concentration of 25mM with increasing time from 6 to 24h. In this concentration (25mM), TiO₂ treatment, up-regulated the *COST* and *TpGAS* in contrast, down-regulated the *TpCarS* with increasing time from 6 to 24h. Salinity treatment affected the expression of all three genes, so that with increasing time, the expression of all three genes was elevated. In conclusion, according to above and HPLC results, it was shown the nanoparticles and salinity treatments can increase parthenolide synthesis in whole plant of Feverfew and then they can be used as elicitor for more production of the metabolite.

Keywords. *Tanacetum parthenium* L., Nanoparticle, SiO₂, TiO₂, Salinity stress, Gene expression analysis.

Abbreviations: *GAS*, Germacrene A Synthase; *COST*, Costunolide Synthase; *CarS*, Caryophyllene Synthase; PTL, Parthenolide; NP, Nanoparticle.

INTRODUCTION

Medicinal plants have a specific role in treatment and prevention of many human diseases. These plants are attracting more attention for produc-

ing safer medicine because it is believed that they rarely have side effects compared to chemical drugs. Feverfew (*Tanacetum parthenium* L.) (Asteraceae), is a diploid ($2n=2x=18$) and perennial medicinal plant. This plant has medical applications on a wide range of disease such as migraine headaches, stomach aches, toothaches, insect bites, rheumatoid arthritis and infertility (Pareek et al. 2011). These medicinal properties are due to the existence of chemical compounds which called secondary metabolites. Terpenoids are the largest class of plant secondary metabolites (Croteau et al. 2000). Sesquiterpene lactones are the main group of terpenoids and frequently are derived from Mevalonic acid (MVA) pathway (Van Klink et al. 2003). *T. parthenium* (L.), contains many sesquiterpene lactones and parthenolide has the most concentration (comprises up to 85%) among the total sesquiterpenes (Pareek et al. 2011).

Parthenolide is used for the treatment of migraine and shows specifically anticancer and anti-inflammatory activity, as well (Tassorelli et al. 2005; Walsh et al. 2011; Mathema et al. 2012; Al-Fatlawi et al. 2015; Wang and Li, 2015). β -caryophyllene is another sesquiterpene lactone distributed in the essential oil of various plants. This compound has represented several biological activities, such as anti-inflammatory, antibiotic, antioxidant (Legault and Pichette, 2007), anticancer (Tundis et al. 2009) and antiproliferative activity (Amiel et al. 2012).

Sesquiterpenes, like parthenolide and β -caryophyllene are synthesized via farnesyl diphosphate (FPP) in Mevalonate pathway. *TpGAS* and *COST* are two genes involved in parthenolide production. At the first step, *TpGAS* converts farnesyl diphosphate to germacrene A, *COST* converts germacrene A acid to costunolide, and parthenolide is one of the derivatives of costunolide. It has been revealed that costunolide synthase is a cytochrom P450 enzyme. *TpCars* is another sesquiterpene synthase in Feverfew which is responsible for the production of β -caryophyllene. This enzyme converts farnesyl diphosphate to β -caryophyllene directly. (Majdi et al. 2011; Liu et al. 2011; Menin et al. 2012; Basha et al. 2016).

Several factors affect the production of secondary metabolites, and elicitors are one of the most efficient ones (Zhao et al. 2005). Plants and plant cells (*in vitro* culture) show the morphological, biochemical and physiological reactions to biological, chemical or physical factors, which is considered as "elicitors." In fact, Elicitation is an induced or enhanced synthesis process of secondary metabolites by the plants and it is a way to ensure their survival, persistence, and competitiveness (Karuppusamy, 2009; Kiong et al. 2005). Elicitors are biological or nonbiological agents that cause biosynthe-

sis and accumulation of secondary metabolites in plants through induction of defense responses (Ramirez-Estrada et al. 2016). The components of microbial cells and poly and oligosaccharides, chemicals such pesticides, heavy metals, and the signaling compounds in plant defense responses (growth factors, e.g. jasmonate) and physical factors such as hyperosmotic stress, UV, cold shock, ultrasound, and pulsed electric field can act as stimulators for hyperproduction of secondary metabolites (Zhao et al. 2010; Gueven and Knorr, 2011; Lin and Wu, 2002).

Nanoparticles (NPs) are new materials which show unique properties related to their physical size. The nanoparticles used in present work are SiO_2 and TiO_2 which are among the most used nanomaterials (Servin et al. 2012; Siddiqui and Al-Whaibi 2014). Titania (TiO_2) has a wide range of applications such as cosmetics (Anselmann, 2001), cancer treatment (Kalbacova et al. 2008), sunscreens or food (Lan et al. 2013). Silica (SiO_2) is another popular metal oxide NPs used in multiple varieties of applications such as disease labeling, drug delivery, photodynamic therapy (Ohulchanskyy et al. 2007), cancer therapy (Cheng et al. 2010; Rosenholm et al. 2010), fertilizer and pest control (Sakr, 2017; Tripathi et al. 2014; Laing et al. 2006).

The effects of TiO_2 NPs on different biological characters of the plant have been done in several studies. Among them, it could be pointed out to the effect of TiO_2 on the growth and microRNA expression profile of tobacco (Frazier et al. 2014), effects of nano- TiO_2 on seed germination (Castiglione et al. 2011), development and mitosis of root tip cells of *Vicia narbonensis* L. and *Zea mays* L. (Castiglione et al. 2011), The effect of SiO_2 and TiO_2 nanoparticles on the expression of GPPS gene (involved in thymoquinone biosynthetic pathway) in *Nigella sativa* L. (Kahila et al. 2017) and finally, Mandeh et al. (2012) which studied *in vitro* influences of TiO_2 nanoparticles on barley tissue culture and quantitative and qualitative characteristics of calli were analyzed after each subculture. In the field of agriculture, SiO_2 has an inhibition effect on the pest (HongShu et al. 2009), Carriers in drug delivery and absorption and transport of nutrients in plants (Liu et al. 2006). The role of nano- SiO_2 in the characteristics of seed germination of tomato has also investigated (Siddiqui and Al-Whaibi, 2014).

Salinity stress is a high concentration of soluble salts in soil and water. Most common soil salinity is caused by high sodium (Na^+) and chloride (Cl^-) (Tavakoli et al. 2010). Salt stress as an environmental factor is another factor that influences on gene expression (Siddiqui and Al-Whaibi 2014).

The effect of salinity on secondary metabolites in plants is various. There are different examples in this case. For example, during NaCl stress in *Swertia chirata*, significant increases ($p \leq 0.05$) was occurred in secondary metabolites at 50mM and initial increase in 100mM NaCl, which falls back to normal levels at the seventh day (Abrol et al. 2012).

In the present work, to evaluate the elicitation role of nanoparticles and abiotic stress on hyperproduction of parthenolide, it was investigated the effect of two NPs (TiO₂ and SiO₂) and salinity stress on the expression of three related genes, *TpGAS*, *COST* and *TpCarS* that are involved in the biosynthetic pathway of PTL and β -caryophyllene in Feverfew.

MATERIALS AND METHODS

Plant material and growth conditions

The seeds of Feverfew were provided by the medicinal plant institute of Shahid Beheshti University, Tehran, Iran. Germinated seeds in pots were grown under controlled circumstances with a 16:8h photoperiod light/dark, 25/18 °C for day/night and supplied with a photosynthetic photon flux density of 3000 lux (Fig. 1).

Nanoparticles characterization

The SiO₂ nanoparticles were purchased from TECNAN Inc. (Tecnología Navarra de Nanoproductos S.L., Spain). A size of 10–15 nm for NPs was estimated (Fig. 2). The XRD measurement clearly showed that the SiO₂ NPs were amorphous. The elemental analysis of the nano-powder by ICP-MS technique (Thermo Elemental VG PQ-ExCell) showed a purity of 99.999%.

The TiO₂ nanoparticles were provided from Degussa Inc., Germany. The analyzed data from XRD showed ~25nm diameter and specific area equal to 55 m²g⁻¹ for the nanoparticles (Ave. of 24.5 nm in diameter, a mixture of anatase and rutile with more proportion of anatase (89.2 %)).

Plant treatment with NPs and Salt

The SiO₂ and TiO₂ NPs, were separately prepared in two concentrations (25 and 50 mg/l) and the solution was used for watering of each pot. For each pot, 80 ml of 0.3M NaCl solution was applied for doing salinity stress treatment. All tissues were collected at the certain times (6, 24 and 48h after irrigation) for gene expression



Fig. 1. The grown plants in pots in green house under controlled condition.

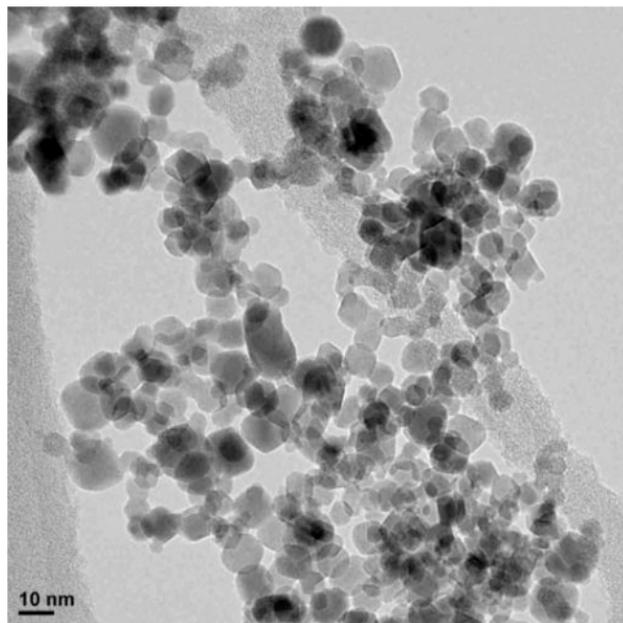


Fig. 2. The TEM micrograph of SiO₂ Nanoparticles.

analysis and phytochemical studies and were instantly flash frozen in liquid nitrogen then stored at -80°C until RNA extraction.

HPLC analysis

The chromatography assay was performed on a 25 cm×4.6 mm with pre-column, Eurospher 100-5 C18 ana-

lytical column provided by KNAUER (Berline, Germany) reversed phase matrix (5 μ m) (Waters) and elution was carried out in a gradient system with acetonitrile as the organic phase (solvent A) and distilled water (solvent B) with the flow-rate of 1 mL min⁻¹. The Peaks were monitored at 220 nm wavelength. Injection volume was 20 μ L and the temperature was maintained at 25°C. All injections were repeated three times (n=3). Calibration graphs were plotted subsequently for linear regression analysis of the peak area with concentration 1, 10, 25, 50, 80, 120, 150 and 200 mg L⁻¹.

RNA extraction

Total RNA was isolated from the leaf tissue by using the Isolation total RNA Kit (Denazist Asia Inc., Mashhad, Iran), according to the manufacturer's instructions. The quantity and quality of extracted RNA were determined respectively by using a spectrophotometer instrument (NanoDrop2000c, Thermo scientific, USA) and Agarose gel electrophoresis. RNA samples were stored at -80 °C until further analysis.

Gene selection and primer designing

In the present work, *TpGAS* and *Cost* genes that involved in biosynthesis of PTL and *TpCarS* as mediator in biosynthesis of β -caryophyllen at Mevalonate pathway in Feverfew (Majdi et al. 2011), plus β -Actin as a house keeping gene were selected and their sequences retrieved from the gene bank, NCBI (<http://www.ncbi.nih.gov>). Four pairs of primers including β -Actin (-5'- AGCATGGTATTGTGAGCAACT-3', R-5'- TGG-GTCATCTTCTCTCTGTTAGC-3'), *TpGAS* (F-5'-TAC-CAGTTTGAGCGTGAAAGA- 3', R- 5'-CAATCAT-GATCTTGAGCTCGT- 3'), *TpCarS* (F-5'-GAGCAT-GTCCACAAAGTATTTAC-3', R-5'- GCATCG-GAATATCTTTACACACAG-3') and *Cost* (F- 5'- GAG-ACACAAGAAGAAGTGAGATCAG-3', R- 5'- AAAG-GTGTAGGAGCATGTAACCTC-3') were designed using primer 3 free software (http://biotools.umassmed.edu/bioapps/primer3_www.cgi).

Primers confirmation was performed by three criteria, including BLAST search in Gene Bank, single peak in qPCR and single band on gel electrophoresis.

cDNA synthesis

cDNA synthesis was performed by Revert Aid First strand cDNA synthesis Kit (Thermo Scientific, USA)

with 200 U of M-MLV RT enzyme, oligo-dT and random Hexamer primers according to manufacturer's instruction.

Quantitative PCR

Quantification of *COST*, *TpGAS* and *TpCarS* genes expression levels in the samples were measured by qPCR using Hot Firepol EvaGreen qPCR master mix (solis BioDyne, Estonia). qPCR was performed according to manufacturer's instruction in Qiagen Real-time PCR System (Rotor-Gene Q, Germany) using above primers. It should be mentioned that before qPCR, the specificity of all primers and optimization of PCR reactions was done with conventional PCR. The relative expression levels were calculated according to Pfaffl method (Pfaffl 2001).

Data Statistics

Expression levels were calculated from the Ct values obtained from triplicate biological samples. Statistical significance analysis of relative gene expression level compared with the reference gene (β -Actin) was performed with completely randomized design (CRD). Mean values of relative expression levels were compared with LSD test (P=0.05) using SPSS ver. 21.0 software.

RESULTS

In this study, the effects of TiO₂ and SiO₂ nanoparticles and salinity stress on expression of *TpGAS*, and *COST* genes that are involved in the biosynthetic pathway of PTL in Feverfew (Fig. 3) were investigated. Also change of *TpCarS* gene expression which produces β -caryophyllen was analyzed. The present study focuses on elicitation effects of nanoparticles and salinity on parthenolide and β -caryophyllen productions in Feverfew.

Gene Expression Analysis

COST Gene

Two concentrations (25 and 50mM) of SiO₂ NP, in two periods of time (6 and 24h) were used. The results showed that, in both 25 and 50mM concentrations, the expression of *COST* gene was enhanced with increasing of time. While in both times, the gene expression in 25mM, was more than 50mM treatment; furthermore, the

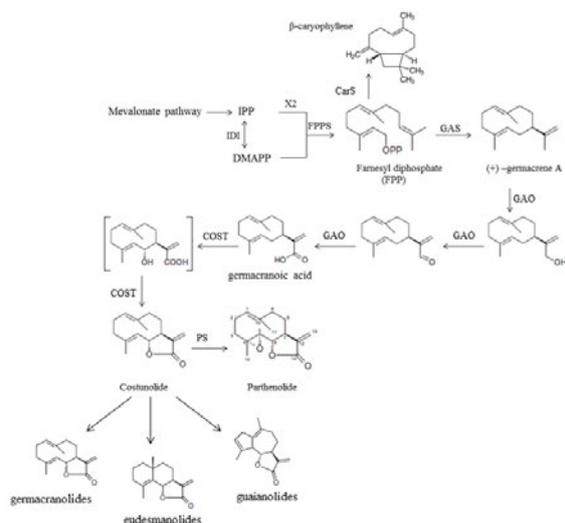


Fig. 3. The suggested biosynthetic pathway for parthenolide and β -caryophyllene synthesis in feverfew (Majdi et al., 2011). CarS: caryophyllene synthase; GAS: germacrene A synthase; GAO: germacrene A oxidase; COST: costunolide synthase; PS: Parthenolide synthase.

expression of the gene in 25mM and after 24h was very high and more than 7-folds in comparison to 6h (Fig. 4a).

The result of TiO₂ NP was similar to SiO₂ treatment at 25mM concentration and increased the expression of COST gene. In 25mM Concentration of TiO₂ NP with increasing time from 6h to 24h, elevation of the gene expression by more than 6-fold (Fig.4b) was observed. In opposite, the gene expression in 50mM of TiO₂ NP was decreased by rising time until close to zero.

As Fig. 4 (a and b) shows, Although the ratio of expression level for SiO₂ and TiO₂ nanoparticles in 6h/24h is nearly the same, but the absolute expression of the COST gene in TiO₂ treated samples (133.8 fold) is significantly more than SiO₂ treated plants (20.2 fold).

The effect of salinity treatment on expression of COST gene were also measured; after 6h and 48h at 0.3 M concentration A gradual increasing trend (2-folds) in gene expression was observed (Fig. 4c).

TpGAS Gene

TpGAS is a gene involved in the parthenolide biosynthesis pathway. The expression analysis of the gene transcript pointed that, it is affected by SiO₂ and TiO₂ NPs and salt stress treatment. The results represented that SiO₂ NPs in 25mM concentration at 24h, significantly increased the expression of GAS compared to 6h treatment (24.5 fold). In opposite, the TiO₂ NPs treatment didn't have a significant effect on the expression of

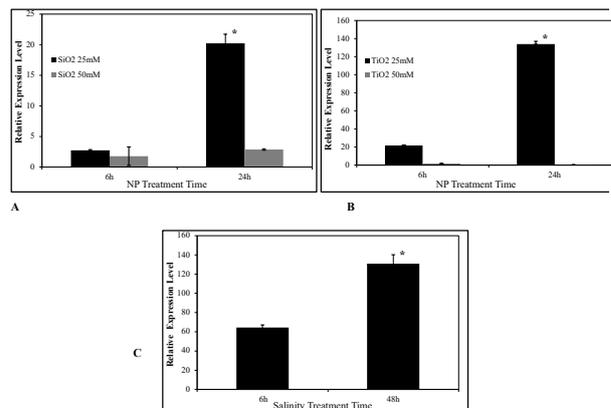


Fig. 4. The effect of elicitors on expression of *COST* gene in feverfew: A) SiO₂ NP treatment; B) TiO₂ NP treatment; C) salinity stress.

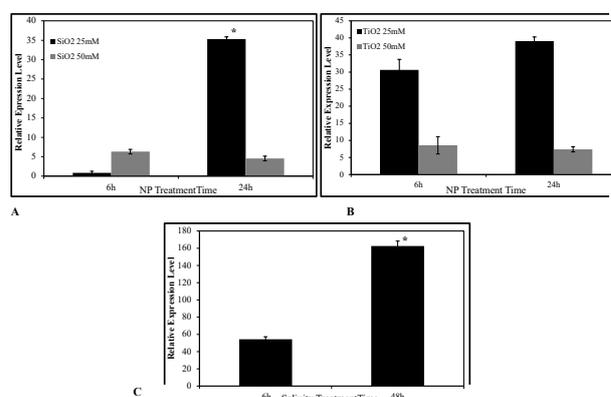


Fig. 5. The effect of elicitors on expression of *TpGAS* gene in feverfew. A: SiO₂ NP treatment; B: TiO₂ NP treatment; C: Salinity stress.

the gene in both time and concentrations (Fig. 5a and b).

The effect of salinity stress was investigated with the same concentration used above. As shown in the Fig.5c, the Expression of *TpGAS*, was statistically enhanced in 48h in comparison to 6h treatment, (~3 fold).

TpCarS Gene

TpCarS is another gene which was analyzed in our experiment. The results of expression analysis showed that the gene was affected by SiO₂ and TiO₂ NPs and salinity treatment, as well. As it can be seen in the Fig. 6a and b, an increase (1.4 fold) and decrease (6 fold) in expression of *TpCarS* in 25mM of SiO₂ and TiO₂ NPs treatment were observed respectively, from 6h to 24h. In contrast, in the 50mM concentration of NPs, no significant changes were detected during the time period.

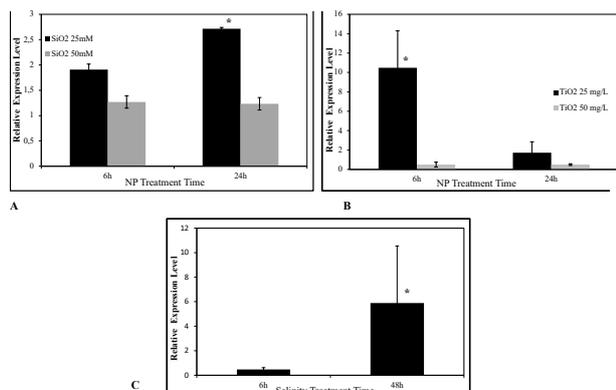


Fig. 6. The effect of elicitors on expression of *TpCarS* gene in feverfew. A: SiO₂ NP treatment; B: TiO₂ NP treatment; C: Salinity stress.

The effect of salinity stress as an environmental elicitor with 300mM of NaCl in two times, including 6h and 48h was investigated. As shown in Fig. 6c, the 48h salt stress elevated the expression of *TpCarS* gene significantly (~12 fold) compared to 6h and 24h treatments.

Parthenolide concentrations in feverfew under different treatments

To assay the parthenolide concentration in different treatments, leaves tissue were analyzed by HPLC (Fig. 7). There were significant differences between parthenolide concentrations ($P < 0.05$) in different treatments (Fig. 8).

The highest and least amount of parthenolide was observed in 25mM of TiO₂ after 24h and control plants with 378.61 μ g/mg and 136.02 μ g/mg, respectively. All treatments increased the concentration of parthenolide in Feverfew leaves compared to control plants ($P < 0.05$). The SiO₂ (25mM) after 6 and 24h treatments increased

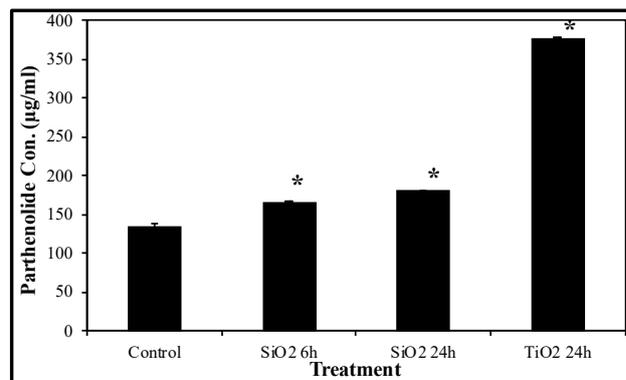


Fig. 8. Parthenolide concentration in *Tanacetum parthenium* leaves at different treatments.

the leaves parthenolide content by 1.23 and 1.37 fold compared to control. The TiO₂ (25mM, 24h) raised the parthenolide amount in leaves far more than SiO₂ NPs with 2.78 fold.

DISCUSSION

Nanoparticles are a new class of man made materials which are emerging in these years. In hence, a new trend in biological sciences is toward investigation of interaction of the synthetic agents to organisms including plants. There are many reports which study the effect of them (TiO₂ and SiO₂) on plants in different kinds of morphological and molecular levels (Siddiqui and Al-Whaibi 2014; Castiglione et al. 2011; Frazier et al. 2014; Kahila et al. 2017), as well *in vivo* culture (Mandeh et al. 2012). A number of studies also prove the key role of different elicitors, including chemical compounds (Van Fürden et al. 2005; Esmailzadeh Bahabadi et al. 2011; Esmailzadeh Bahabadi et al. 2014;

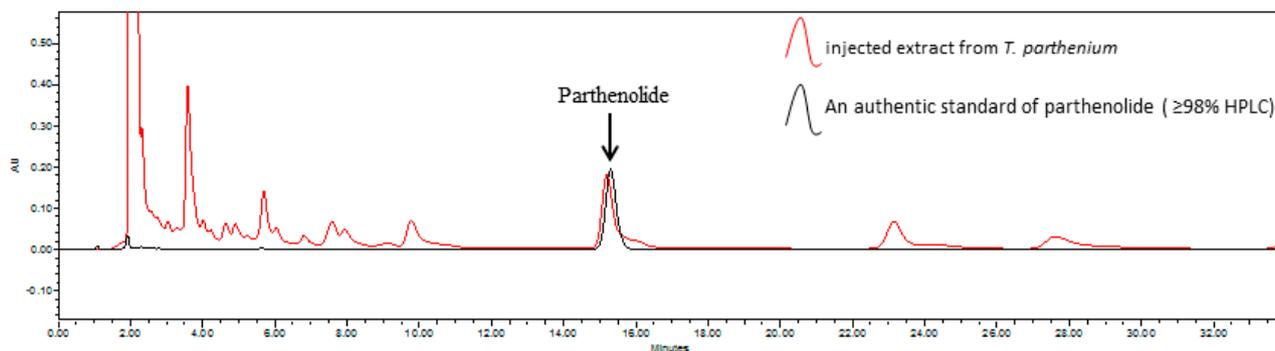


Fig. 7. The HPLC analysis of extract from *T. parthenium* and 110 ppm of standard parthenolide from Sigma-Aldrich company (USA). Minutes: Run time, AU: Absorption intensity.

Wang et al. 2007; Marsh et al. 2014), biotic (Kim et al. 2001; Jeong et al. 2005; Savitha et al. 2006; Wu et al. 2007; Kang et al. 2009; Gao et al. 2011; Swaroopa et al. 2013; Ahmed and Baig. 2014) and abiotic (Akula and Ravishankar 2011; Chan et al. 2010; Szakiel et al. 2011) on metabolite production in different plants. However, nearly most of studies have been conducted on cell or hairy root culture and investigation on *in vivo* whole plant elicitation is limit. The aim of the present work was to determine the elicitation role of nanoparticles and salt stress on metabolite hyperproduction through gene expression in whole plant.

NPs and salinity treatments affect the parthenolide biosynthesis genes

The effect of different kind of elicitors was investigated to find a clear view about the impact of elicitors on the expression of genes involved in the secondary metabolites biosynthesis pathway in *T. parthenium*. It seems that SiO₂ NPs in low concentration can have a positive effect on parthenolide production in Feverfew. In opposite to SiO₂, TiO₂ NPs treatment created a different expression pattern of the *TpGAS*, *COST* and *TpCarS* genes.

Dimkpa et al. (2012) demonstrated the beneficial effects of CuO and ZnO NPs on IAA production *in vitro* in the soil isolate *P. chlororaphis* O6. An increased IAA production with exposure to sublethal levels CuO NPs was observed in their study. They explained an ion release and nonspecific mechanism for increased level of IAA due to CuO and ZnO NPs treatment, respectively.

The salt stress up-regulated all the mentioned genes in our study. It seems, this is an evolutionary mechanism in medicinal plant to tolerance abiotic stress with producing of metabolites including PTL compound in Feverfew.

A comparison between the salts stress and the nanoparticles effect on the genes and hyperproduction of parthenolide, mentioned that, the NPs are more effective than salt stress, because they affected two key genes, including *TpGAS* and *COST* which catalyzes two steps of parthenolide biosynthesis pathway compared to salt stress which affect only on *TpCarS* gene.

The drought and salt stresses cause common reactions in plants. Cellular dehydration then osmotic stress and transfer of water from the cytoplasm to vacuoles are a result of both stresses (Akula and Ravishankar, 2011).

Salinity stress often causes both ionic and osmotic stress in plants, which increases or decreases specific secondary metabolites in plants (Mahajan and Tuteja, 2005). For example, Parida and Das (2005) found

that anthocyanins are increased in response to salt stress. Oppositely, Daneshmand et al. (2010) demonstrated that salinity stress decreases the anthocyanin level in the salt-sensitive species. As the Plant polyamines are involved in plant response to salinity, changes of free and bound polyamines levels has been reported in sunflower (*Helianthus annuus* L.) roots under salinity stress (Mutlu and Bozcuk, 2007). The endogenous JA (Jasmonic acid) under salt stress in tomato cultivars has been found (Pedranzani et al. 2003). Polyphenols are another group of metabolites which their synthesis and accumulation is usually is stimulated as a reaction to biotic or abiotic stresses (Dixon and Paiva, 1995; Muthukumarasamy et al. 2000). In some of plants such red peppers (Navarro et al. 2006); a raise in polyphenol content with increasing level of salt in different tissues has also been reported (Parida and Das, 2005).

Positive correlation between TpGAS and COST expression and parthenolide concentration under NPs Treatments

A positive correlation between *TpGAS* and *COST* expression and parthenolide concentration was observed in SiO₂ and TiO₂ treatments (25mM and 24h) compared to control plants, in contrast to *TpCarS* of which its expression had a negative correlation with increasing treatment time. Although few studies have revealed the genotoxic potential of TiO₂ nanoparticles in the plant systems (*A. cepa* and *N. tabacum*) with DNA damaging effect (Ghosh et al. 2010), in this work, TiO₂ nanoparticles had a positive effect on parthenolide production. *TpGAS* encodes the enzyme that highly likely catalyzes the first step in parthenolide biosynthesis, germacrene A synthase (Fig.3) (Majdi et al. 2011). Then, it seems that the Feverfew plant responds to the NPs by up-regulation of the responsive genes and then metabolite production.

TpCarS (β -Caryophyllene synthase) encodes β -Caryophyllene which is an anti-inflammatory (Martin et al. 1993; Tambe et al. 1996) and anti-carcinogenic (Kubo et al. 1996; Zheng et al. 1992) compound and a common and quite widely distributed sesquiterpene in plants (Knudsen et al. 1993; Kubo et al. 1996).

Similar to *Artemisia annua* plant (Chen et al. 2011), in *Tanacetum parthenium* (Bouwmeester et al. 2002) farnesyl diphosphate (FPP) (Fig.3) is an initial point and serves as a basic precursor to synthesize the various classes of sesquiterpenes with divergent structures and functions by different synthases such *TpCarS* through competitive pathways. Therefore, depending on their competition with the available FPP pool, the critical step catalyzed by different sesquiterpene synthases shows a metabolic regulating to direct the cellular carbon flux

towards parthenolide or other sesquiterpenes. Therefore, it seems, the nanoparticles directly and exclusively induce the genes toward more production of parthenolide compared to β -Caryophyllene.

Different studies have proved hyperproduction of secondary metabolite related to use of elicitors, including Jasmonate (Walker et al. 2002; Zhao et al. 2010; Tocci et al. 2012; Tocci et al. 2011; Gadzovska et al. 2013; Cui et al. 2014). It has been found that Jasmonic acid (JA) and its methyl esters, methyl jasmonate (MeJA), are important signaling compounds in the process of elicitation leading to the hyper production of various secondary metabolites (Walker et al. 2002). This compound plays a key role in signal transduction processes involved in defense responses in plant and has shown that is effective to induce the production of secondary metabolites in cell cultures (Walker et al. 2002; Zhao et al. 2010; Tocci et al. 2012; Tocci et al. 2011; Gadzovska et al. 2013; Cui et al. 2014).

The effect of the elicitors on flavonoid production was reported by Wang et al. (2015). They showed that the Flavonoid content is promoted by MeJA and SA induction, which were 2.1 and 1.5 times higher in comparison to control cultures, respectively.

A list of reports on different plant species have demonstrated a positive and strong correlation between terpene content and the level of the related mRNA transcripts, which proves the terpenoid biosynthesis is majorly regulated at the transcript level (Nagegowda, 2010).

CONCLUSION

The results of our work, showed a significant effect of nanoparticles in low concentration and salinity stress on the expression of two genes involved in parthenolide biosynthesis pathway, also *TpCarS* as a caryophyllen synthase in Feverfew. It seems due to the positive effect of SiO₂ NP compared to TiO₂, on the expression of all key genes for parthenolide and β -caryophyllen productions in Feverfew, these nanoparticles can be used as efficient elicitors and useful additives to soil for increasing of secondary metabolite production in the whole plant, practically. As it was mentioned above, the metabolites are used as medicinal compounds for patients with migraine disease and different type of cancers; in hence, our results can suggest a simple and cost effective technique for mass production of them. However, due to a dominant view among researchers about the destructive role of nanoparticles in the environment, it is necessary for more research for investigation about side effect of

the nanoparticles on plant growth and development in future. In conclusion, in this work, it was tried to explain the role of nanoparticle by this point of view which the nanomaterial, can also be valuable for more providing of human medicinal necessities from herbs and then, it opened a new window toward nanoparticle application for medicinal plants studies. it should be mentioned that, in this project the short-term effects of elicitors on secondary metabolite production were investigated and further studies are needed to determine the long-term impact of these elicitors on plants gene expression.

ACKNOWLEDGEMENTS

We would to thank the University of Tehran for instrumental supports and all people who help us in this work.

AUTHOR CONTRIBUTION STATEMENT

MR conceived and designed research. MK conducted experiments. MR contributed new reagents or analytical tools. MR analyzed data. MK, MR and AE wrote the manuscript. All authors read and approved the manuscript.

AVAILABILITY OF DATA AND MATERIALS

The datasets during and/or analyzed during the current study available from the corresponding author on reasonable request.

REFERENCES

- Abrol E, Vyas D, Koul S. 2012. Metabolic shift from secondary metabolite production to induction of anti-oxidative enzymes during NaCl stress in *Swertia chirata* Buch.-Ham. *Acta Physiol Plant.* 34(2):541-546.
- Ahmed SA, Baig MM. 2014. Biotic elicitor enhanced production of psoralen in suspension cultures of *Psoralea corylifolia* L. *Saudi J Biol Sci.* 21(5):499-504.
- Akula R, Ravishankar GA. 2011. Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signal Behav.* 6(11):1720-1731.
- Al-Fatlawi AA, Al-Fatlawi AA, Irshad M, Rahisuddin, Ahmad A. 2015. Effect of parthenolide on growth and apoptosis regulatory genes of human cancer cell lines. *Pharm Biol.* 53(1):104-109.

- Amiel E, Ofir R, Dudai N, Soloway E, Rabinsky T, Rachmilevitch S. 2012. β -Caryophyllene, a compound isolated from the biblical balm of gilead (*Commiphora gileadensis*), is a selective apoptosis inducer for tumor cell lines. *Evid Based Complement Alternat Med*. 2012: 1-8.
- Anselmann R. 2001. Nanoparticles and nanolayers in commercial applications. *J Nanopart Res*. 3(4): 329-336.
- Basha RH, Sankaranarayanan C. 2016. β -Caryophyllene, a natural sesquiterpene lactone attenuates hyperglycemia mediated oxidative and inflammatory stress in experimental diabetic rats. *Chem Biol Interac*. 245: 50-58.
- Bouwmeester HJ, Kodde J, Verstappen FWA, Altug IG, de Kraker JW, Wallaart TE. 2002. Isolation and characterization of two germacrene A synthase cDNA clones from chicory. *Plant Physiol*. 129(1):134-144.
- Castiglione MR, Giorgetti L, Geri C, Cremonini R. 2011. The effects of nano-TiO₂ on seed germination, development and mitosis of root tip cells of *Vicia narbonensis* L. and *Zea mays* L. *J. Nanopart Res*. 13(6):2443-2449.
- Chan LK, Koay SS, Boey PL, Bhatt A. 2010. Effects of abiotic stress on biomass and anthocyanin production in cell cultures of *Melastoma malabathricum*. *Biol Res*. 43:127-135.
- Chen JL, Fang HM, Ji YP, Pu GB, Guo YW, Huang LL, Du ZG, Liu BY, Ye HC, Li GF, Wang H. 2011. Artemisinin biosynthesis enhancement in transgenic *Artemisia annua* plants by downregulation of the β -caryophyllene synthase gene. *Planta medica*. 77(15):1759-1765.
- Cheng SH, Lee CH, Chen MC, Souris JS, Tseng FG, Yang CS, ... & Lo LW. 2010. Tri-functionalization of mesoporous silica nanoparticles for comprehensive cancer theranostics—the trio of imaging, targeting and therapy. *J. Mater. Chem*. 20: 6149-6157
- Croteau R, Kutchan TM, Lewis NG. 2000. Natural products (secondary metabolites). *Biochem Mol Biol Plants*. 24: 1250-1319.
- Cui XH, Murthy HN, Paek KY. 2014. Pilot-scale culture of *Hypericum Perforatum* L. adventitious roots in airlift bioreactors for the production of bioactive compounds. *Appl Biochem Biotechnol*. 174(2):784-792.
- Daneshmand F, Arvin MJ, Kalantari KM. 2010. Physiological responses to NaCl stress in three wild species of potato in vitro. *Acta Physiol Plant*. 32(1):91-101.
- Dimkpa CO, Zeng J, McLean JE, Britt DW, Zhan J, Anderson AJ. 2012. Production of indole-3-acetic acid via the indole-3-acetamide pathway in the plant-beneficial bacterium *Pseudomonas chlororaphis* O6 is inhibited by ZnO nanoparticles but enhanced by CuO nanoparticles. *Appl Environ Microbiol*. 78(5):1404-1410.
- Dixon RA, Paiva NL. 1995. Stress-induced phenylpropanoid metabolism. *The plant cell*. 7(7):1085-1097.
- Esmailzadeh Bahabadi S, Sharifi M, Murata J, Satake H. 2014. The effect of chitosan and chitin oligomers on gene expression and lignans production in *Linum album* cell cultures. *J Med Plants*. 1(49):46-53.
- Esmailzadeh Bahabadi S, Sharifi M, Safaie N, Murata J, Yamagaki T, Satake H. 2011. Increased lignan biosynthesis in the suspension cultures of *Linum album* by fungal extracts. *Plant Biotechnol Rep*. 5(4):367-373.
- Frazier TP, Burklew CE, Zhang B. 2014. Titanium dioxide nanoparticles affect the growth and microRNA expression of tobacco (*Nicotiana tabacum*). *Funct Integr Genomic*. 14(1):75-83.
- Gadzovska S, Maury S, Delaunay A, Spasenoski M, Hagege D, Courtois D, Joseph C. 2013. The influence of salicylic acid elicitation of shoots, callus, and cell suspension cultures on production of naphthodianthrones and phenylpropanoids in *Hypericum perforatum* L. *Plant Cell Tiss Organ Cult*. 113(1):25-39.
- Gao FK, Yong YH, Dai CC. 2011. Effects of endophytic fungal elicitor on two kinds of terpenoids production and physiological indexes in *Euphorbia pekinensis* suspension cells. *J Med Plants Res*. 5(18):4418-4425.
- Ghosh M, Bandyopadhyay M, Mukherjee A. 2010. Genotoxicity of titanium dioxide (TiO₂) nanoparticles at two trophic levels: plant and human lymphocytes. *Chemosphere*. 81(10):1253-1262.
- Gueven A, Knorr D. 2011. Isoflavonoid production by soy plant callus suspension culture. *J Food Eng*. 103(3):237-243.
- HongShu X, HaiBo C, JunBo L, JingHua L, Chen G. 2009. Inhibition effect of SiO₂ hydrosol against leaf mold of tomato. *J Jilin Agric Uni*. 31(3):252-254.
- Jeong GT, Park DH, Ryu HW, Hwang B, Woo JC, Kim D, Kim SW. 2005. Production of antioxidant compounds by culture of *Panax ginseng* CA Meyer hairy roots. In: *Twenty-Sixth Symposium on Biotechnology for Fuels and Chemicals* pp 1147-1157. Humana Press.
- Kahila MMH, Najy AM, Rahaie M, Mir-Derikvand M. 2017. Effect of Nanoparticle Treatment on Expression of a key gene involved in Thymoquinone biosynthetic pathway in *Nigella sativa* L. *Nat Prod Res*. 28: 1-5.
- Kang SM, Min JY, Kim YD, Karigar CS, Kim SW, Goo GH, Choi MS. 2009. Effect of biotic elicitors on the

- accumulation of bilobalide and ginkgolides in *Ginkgo biloba* cell cultures. *J Biotechnol.* 139(1):84-88.
- Kalbacova M, Macak JM, Schmidt-Stein F, Mierke CT, Schmuki P. 2008. TiO₂ nanotubes: photocatalyst for cancer cell killing. *Phys Status Solidi R.* 2(4): 194-196
- Karuppusamy S. 2009. A review on trends in production of secondary metabolites from higher plants by in vitro tissue, organ and cell cultures. *J Med Plants Res.* 3(13):1222-1239.
- Kim CY, Im HW, Kim HK, Huh H. 2001. Accumulation of 2, 5-dimethoxy-1, 4-benzoquinone in suspension cultures of *Panax ginseng* by a fungal elicitor preparation and a yeast elicitor preparation. *Appl Microbiol Biotechnol.* 56(1):239-242.
- Kiong AL, Mahmood M, Fadzillah NM, Daud SK. 2005. Effects of precursor supplementation on the production of triterpenes by *Centella asiatica* callus culture. *Pak J Biol Sci.* 8(8):1160-1169.
- Knudsen JT, Tollsten L, Bergstrom LG. 1993. Floral scents—a checklist of volatile compounds isolated by head-space techniques. *Phytochemistry.* 33(2):253-280.
- Kubo I, Chaudhuri SK, Kubo Y, Sanchez Y, Ogura T, Saito T, Ishikawa H, Haraguchi H. 1996. Cytotoxic and antioxidant sesquiterpenoids from *Heterotheca inuloides*. *Planta Med.* 62(5):427-430.
- Laing MD, Gatarayihya MC, Adandonon A. 2006. Silicon use for pest control in agriculture: a review. *Proceedings of the South African Sugar Technologists' Association.* 80: 278-286.
- Lan Y, Lu Y and Ren Z. 2013. Mini review on photocatalysis of titanium dioxide nanoparticles and their solar applications. *Nano Energy.* 2(5): 1031-1045.
- Legault J, Pichette A. 2007. Potentiating effect of β -caryophyllene on anticancer activity of α -humulene, isocaryophyllene and paclitaxel. *J Pharm Pharmacol.* 59(12): 1643-1647.
- Lin L, Wu J. 2002. Enhancement of shikonin production in single-and two-phase suspension cultures of *Lithospermum erythrorhizon* cells using low-energy ultrasound. *Biotechnol Bioeng.* 78(1):81-88.
- Liu F, Wen LX, Li ZZ, Yu W, Sun HY, Chen JF. 2006. Porous hollow silica nanoparticles as controlled delivery system for water-soluble pesticide. *Mater Res Bull.* 41(12):2268-2275.
- Mahajan S, Tuteja N. 2005. Cold, salinity and drought stresses: An overview. *Arch Biochem Biophys.* 444:139-158.
- Majdi M, Liu Q, Karimzadeh G, Malboobi MA, Beekwilder J, Cankar K, de Vos R, Todorović S, Simonović A, Bouwmeester H. 2011. Biosynthesis and localization of parthenolide in glandular trichomes of feverfew (*Tanacetum parthenium* L. Schulz Bip.). *Phytochemistry.* 72(14):1739-1750.
- Mandeh M, Omidi M, Rahaie M. 2012. In vitro influences of TiO₂ nanoparticles on barley (*Hordeum vulgare* L.) tissue culture. *Biol Trace Elem Res.* 150(1-3):376-380.
- Marsh Z, Yang T, Nopo-Olazabal L, Wu S, Ingle T, Joshee N, Medina-Bolivar F. 2014. Effect of light, methyl jasmonate and cyclodextrin on production of phenolic compounds in hairy root cultures of *Scutellaria lateriflora*. *Phytochemistry.* 107:50-60.
- Martin S, Padilla E, Icete MA, Galvez J, Jimenez J, Zarzelo Z. 1993. Anti-inflammatory activity of the essential oil of *Bupleurum fruticosum*. *Planta Med.* 59:533-536.
- Mathema VB, Koh YS, Thakuri BC, Sillanpää M. 2012. Parthenolide, a sesquiterpene lactone, expresses multiple anti-cancer and anti-inflammatory activities. *Inflammation.* 35(2):560-565.
- Menin B, Comino C, Portis E, Moglia A, Cankar K., Bouwmeester HJ, Lanteri S, Beekwilder J. 2012. Genetic mapping and characterization of the globe artichoke (+)-germacrene A synthase gene, encoding the first dedicated enzyme for biosynthesis of the bitter sesquiterpene lactone cynaropicrin. *Plant Sci.* 190: 1-8.
- Muthukumarasamy M, Gupta SD, Panneerselvam R. 2000. Enhancement of peroxidase, polyphenol oxidase and superoxide dismutase activities by triadimefon in NaCl stressed *Raphanus sativus* L. *Biol Plant.* 43(2):317-320.
- Mutlu F, Bozcuk S. 2007. Salinity-induced changes of free and bound polyamine levels in sunflower (*Helianthus annuus* L.) roots differing in salt tolerance. *Pak J Bot.* 39(4):1097-1102.
- Nagegowda DA. 2010 Plant volatile terpenoid metabolism: biosynthetic genes, transcriptional regulation and subcellular compartmentation. *FEBS let.* 584(14):2965-2973.
- Navarro JM, Flores P, Garrido C, Martinez V. 2006 Changes in the contents of antioxidant compounds in pepper fruits at different ripening stages, as affected by salinity. *Food Chem.* 96(1):66-73.
- Ohulchanskyy TY, Roy I, Goswami LN, Chen Y, Bergey EJ, Pandey RK., Oseroff AR, Prasad PN. 2007. Organically modified silica nanoparticles with covalently incorporated photosensitizer for photodynamic therapy of cancer. *Nano lett.* 7(9): 2835-2842.
- Pareek A, Suthar M, Rathore GS, Bansal V. 2011. Feverfew (*Tanacetum parthenium* L.): A systematic review. *Pharmacogn Rev.* 5(9):103-110.

- Parida AK, Das AB. 2005. Salt tolerance and salinity effects on plants: a review. *Ecotoxicol Environ Saf.* 60(3):324-349.
- Pedranzani H, Racagni G, Alemano S, Miersch O, Ramirez I, Peña-Cortés H, Taleisnik E, Machado-Domenech E, Abdala G. 2003. Salt tolerant tomato plants show increased levels of jasmonic acid. *Plant Growth Regul.* 41(2):149-58.
- Pfaffl, MW. 2001. A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic acids res.* 29(9): 45-45.
- Ramirez-Estrada K, Vidal-Limon H, Hidalgo D, Moyano E, Golenioswki M, Cusidó RM, Palazon J. 2016. Elicitation, an effective strategy for the biotechnological production of bioactive high-added value compounds in plant cell factories. *Molecules.* 21(2):182.
- Rosenholm JM, Sahlgren C, Linden M. 2010. Towards multifunctional, targeted drug delivery systems using mesoporous silica nanoparticles – opportunities & challenges'. *Nanoscale.* 2: 1870-1883
- Sakr N, 2017. The role of silicon (Si) in increasing plant resistance against insect pests review article. *Acta Phytopathol Entomol Hung.* 52(2): 185-204.
- Savitha BC, Thimmaraju R, Bhagyalakshmi N, Ravishankar GA. 2006. Different biotic and abiotic elicitors influence betalain production in hairy root cultures of *Beta vulgaris* in shake-flask and bioreactor. *Process Biochem.* 41(1):50-60.
- Servin AD, Castillo-Michel H, Hernandez-Viezcas JA, Diaz BC, Peralta-Videa JR, Gardea-Torresdey JL. 2012. Synchrotron micro-XRF and micro-XANES confirmation of the uptake and translocation of TiO₂ nanoparticles in cucumber (*Cucumis sativus*) plants. *Environ Sci Technol.* 46(14):7637-7643.
- Siddiqui MH, Al-Whaibi MH. 2014. Role of nano-SiO₂ in germination of tomato (*Lycopersicon esculentum* seeds Mill.). *Saudi J Biol Sci.* 21(1):13-17.
- Swaroop G, Anuradha M, Pullaiah T. 2013. Elicitation of forskolin in suspension cultures of *Coleus forskohlii* (willd.) Briq. using elicitors of fungal origin. *Curr Trends Biotechnol Pharm.* 7(3):755-762.
- Szakiel A, Paczkowski C, Henry M. 2011. Influence of environmental abiotic factors on the content of saponins in plants. *Phytochem Rev.* 10(4): 471-91.
- Tambe Y, Tsujiuchi H, Honda G, Ikeshiro Y, Tanaka S. 1996. Gastric cytoprotection of the non-steroidal anti-inflammatory sesquiterpene, b-caryophyllene. *Planta Med.* (62):469-470.
- Tassorelli C, Greco R, Morazzoni P, Riva A, Sandrini G, Nappi G. 2005. Parthenolide is the component of *tanacetum parthenium* that inhibits nitroglycerin-induced Fos activation: studies in an animal model of migraine. *Cephalalgia.* 25(8):612-621.
- Tavakkoli E, Rengasamy P, McDonald GK. 2010. High concentrations of Na⁺ and Cl⁻ ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. *J Experimental botany.* 61(15): 4449-4459.
- Tocci N, D'Auria FD, Simonetti G, Panella S, Palamara AT, Pasqua G. 2012. A three-step culture system to increase the xanthone production and antifungal activity of *Hypericum perforatum* subsp. *angustifolium* in vitro roots. *Plant Physiol Biochem.* 57:54-58.
- Tocci N, Simonetti G, D'Auria FD, Panella S, Palamara AT, Valletta A, Pasqua G. 2011. Root cultures of *Hypericum perforatum* subsp. *angustifolium* elicited with chitosan and production of xanthone-rich extracts with antifungal activity. *Appl Microbiol Biotechnol.* 91(4):977-987.
- Tripathi DK, Singh VP, Gangwar S, Prasad SM, Maurya JN, Chauhan DK. 2014. Role of silicon in enrichment of plant nutrients and protection from biotic and abiotic stresses, in: *In Improvement of crops in the era of climatic changes.* New York, pp. 39-56.
- Tundis R, Loizzo MR, Bonesi M, Menichini F, Dodaro D, Passalacqua NG, Statti G, Menichini F. 2009. In vitro cytotoxic effects of *Senecio stabianus* Lacaita (Asteraceae) on human cancer cell lines. *Nat Prod Res.* 23(18): 1707-1718.
- Van Fürden B, Humburg A, Fuss E. 2005. Influence of methyl jasmonate on podophyllotoxin and 6-methoxypodophyllotoxin accumulation in *Linum album* cell suspension cultures. *Plant Cell Rep.* 24(5):312-317.
- Van Klink J, Becker H, Andersson S, Boland W. 2003. Biosynthesis of anthecotuloid, an irregular sesquiterpene lactone from *Anthemis cotula* L. (Asteraceae) via a non-farnesyl diphosphate route. *Org Biomol Chem.* 1(9): 1503-1508.
- Walker TS, Bais HP, Vivanco JM. 2002. Jasmonic acid-induced hypericin production in cell suspension cultures of *Hypericum perforatum* L. (St. John's wort). *Phytochemistry.* 60(3):289-293.
- Walsh EL, Ashe S, Walsh JJ. 2011. Nature's Migraine Treatment: Isolation and Structure Elucidation of Parthenolide from *Tanacetum parthenium*. *J Chem Educ.* 89(1):134-137.
- Walsh EL, Ashe S, Walsh JJ. 2011. Nature's Migraine Treatment: Isolation and Structure Elucidation of Parthenolide from *Tanacetum parthenium*. *J Chem Educ.* 89: 134-137.
- Wang J, Qian J, Yao L, Lu Y. 2015. Enhanced production of flavonoids by methyl jasmonate elicitation in cell

- suspension culture of *Hypericum perforatum*. *Biore-sour Bioprocess*. 18;2(1):5.
- Wang M, Li Q. 2015. Parthenolide could become a promising and stable drug with anti-inflammatory effects. *Nat Prod Res*. 29(12):1092-1101.
- Wang YD, Wu JC, Yuan YJ. 2007. Salicylic acid-induced taxol production and isopentenyl pyrophosphate biosynthesis in suspension cultures of *Taxus chinensis* var. *mairei*. *Cell Biol In*. 31(10):1179-1183.
- Wu JY, Ng J, Shi M, Wu SJ. 2007. Enhanced secondary metabolite (tanshinone) production of *Salvia miltiorrhiza* hairy roots in a novel root-bacteria coculture process. *Appl Microbiol Biotechnol*. 77(3):543-550.
- Zhao J, Davis LC, Verpoorte R. 2005. Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnol Adv*. 23(4):283-333.
- Zhao JL, Zhou LG, Wu JY. 2010. Effects of biotic and abiotic elicitors on cell growth and tanshinone accumulation in *Salvia miltiorrhiza* cell cultures. *Appl Microbiol Biotechnol*. 87(1):137-144.
- Zheng GQ, Kenney PM, Lam LK. 1992. Sesquiterpenes from clove (*Eugenia caryophyllata*) as potential anti-carcinogenic agents. *J Nat Prod*. 55(7):999-1003.