



Citation: L. Gonçalves Rodrigues, J. de Senna Pereira, J. Mena Barreto de Freitas, C. Ubessi, S. Bosio Tedesco (2020) Antiproliferative analysis of aqueous extracts of cabreúva (*Myrocarpus frondosus*) on the *Allium cepa* cell cycle. *Caryologia* 73(4): 39-44. doi: 10.13128/caryologia-776

Received: December 15, 2019

Accepted: December 19, 2020

Published: May 19, 2021

Copyright: © 2020 L. Gonçalves Rodrigues, J. de Senna Pereira, J. Mena Barreto de Freitas, C. Ubessi, S. Bosio Tedesco. This is an open access, peer-reviewed article published by Firenze University Press (<http://www.fupress.com/caryologia>) and distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Competing Interests: The Author(s) declare(s) no conflict of interest.

Antiproliferative analysis of aqueous extracts of cabreúva (*Myrocarpus frondosus*) on the *Allium cepa* cell cycle

LUÍSA GONÇALVES RODRIGUES*, JULIA DE SENNA PEREIRA, JÉSSICA MENA BARRETO DE FREITAS, CASSIANE UBESSI, SOLANGE BOSIO TEDESCO

Biology Department, Federal University of Santa Maria, Santa Maria-RS, Brazil

*Corresponding author- E-mail: luisagr.bio@gmail.com

Abstract. *Myrocarpus frondosus* is widely used in Brazilian folk medicine for bronchitis, gastritis, ulcers and wound asepsis. However, there is a scarcity of scientific data proving the safe use of tea of this species or if there is any toxic effect on the human organism. This study aimed to evaluate the antiproliferative effect of aqueous extracts of cabreúva on the *Allium cepa* cell cycle. The aqueous extracts were prepared from leaves, bark and roots (dried material) of the species in two concentrations: 2.5 and 5.0 grams in 250 mL of distilled water. The aqueous extract of the leaves was obtained by infusion and the aqueous extract of the bark and roots by decoction. Distilled water was used as negative control and glyphosate 2% as positive control. Eight groups of four onion bulbs were evaluated and each group corresponded to one treatment and, in each group, 4000 cells were analyzed and the mitotic index was calculated. The results demonstrated reduction of mitotic indices in all treatments when compared to the negative control in water. The aqueous extracts of cabreúva in the studied concentrations have antiproliferative action on the *Allium cepa* cell cycle, ensuring the safe use of tea of this medicinal species.

Keywords: *Myrocarpus frondosus*, medicinal plant, mitotic index.

1. INTRODUCTION

Plants with medicinal potential have been widely used to treat various diseases and are often the only medicine available to the population (Fachinetto and Tedesco, 2009). However, indiscriminate use coupled with lack of knowledge about the medicinal species can cause harm to health (Frescura et al. 2012). The genus *Myrocarpus* is exclusively South American, being *Myrocarpus frondosus* Allemão the only species recorded in southwestern Paraguay, northern Argentina and southern, southeastern and northeastern Brazil (Lorenzi and Matos, 2008).

The species *M. frondosus* belongs to the family Leguminosae, is characterized by being a large tree plant and popularly known as cabreúva (Cabrera et al. 2012). Cabreúva is considered a relevant species because, besides being used as a medicinal plant, it is also used in reforestation of degraded areas

(Scoti et al. 2011; Santi et al. 2017). It is popularly used for diarrhea, gastritis, ulcers, wound aseptis, has expectorant effect, anti-inflammatory and antimicrobial activity (Pereira Junior et al. 2014; Santi et al. 2017). Despite its widespread use in folk medicine, there are no scientific studies to prove its effectiveness and / or rule out possible unwanted or adverse effects from its consumption. Thus, this fact highlights the need for studies that evaluate the action of aqueous extracts of the species *Myrocarpus frondosus* on organisms.

The effects of aqueous extracts can be assessed by testing with bioindicators, which generally include subsystems of a complete organism used to identify a specific target (Leme and Marin-Morales, 2009). From the results obtained, the mitotic index is calculated. Mitotic index values are used as indicative of proper cell proliferation and measured by the *Allium cepa* plant test system. This test has been widely used as a genotoxicity bioindicator (Tedesco and Laghinghouse, 2012). The efficiency of the *Allium cepa* system is due to its proliferation kinetic characteristic, rapid root growth, large number of dividing cells, high tolerance to different cultivation conditions, permanent availability, easy handling, reduced chromosome number ($2n = 16$) and easy viewing under the microscope (Caritá and Marin-Morales, 2008). And from this test it is possible to monitor the effects of medicinal plant extracts, ensuring their safe use by the population in the treatment of diseases and sporadic consumption (Ubessi et al. 2019).

Considering the medicinal importance of the species *Myrocarpus frondosus* and the lack of information regarding its antiproliferative activity, the effect of aqueous extracts from leaves, bark and roots on the *Allium cepa* cell cycle was evaluated.

2. MATERIAL AND METHODS

2.1. Obtaining plant material

The plant material of the species *Myrocarpus frondosus* was collected from a population located in the west of Santa Maria, Rio Grande do Sul, Brazil, under the coordinates 29°41'23.6"S 53°50'45.3"W. The experiment was carried out at the Plant Cytogenetics and Genotoxicity Laboratory at the Federal University of Santa Maria (UFSM).

2.2. Preparation of aqueous extracts

For the preparation of aqueous extracts leaves, bark and roots (dried material) were used in two concentra-

tions: 2.5 and 5.0 grams (g) in 250 mL of distilled water. The leaves were placed in a container containing boiling water, remaining infused for 10 minutes. The extracts of the bark and roots of cabreúva were prepared by decoction in a period of 10 minutes. All extracts after 10 minutes were strained and stored until room temperature.

2.3. *Allium cepa* test

The *Allium cepa* test was developed at the Plant Cytogenetics and Genotoxicity Laboratory (UFSM) and organized into eight groups of four onion bulbs, which were placed for rooting in distilled water for a period of 72 hours. Distilled water was used as negative control and glyphosate 2% as positive control. The evaluated treatments are described in Table 1.

After rooting, the bulbs remained in contact with the treatments described in Table 1 for a period of 24 hours. Time lapse mentioned, the roots were detached from the bulbs and fixed in Carnoy 3:1 (ethanol: acetic acid) for 24 hours at room temperature. Soon after, the roots were placed in 70% ethanol and stored under refrigeration until blades preparation.

2.4. Preparation of blades

Two blades per bulb were made, and 500 cells per blades were analyzed, totaling 4000 cells per treatment. The preparation of the blades was performed according to the crushing technique (Guerra and Souza, 2002). In this procedure the roots were washed in distilled water and hydrolyzed for 5 minutes in HCL 1N at room temperature. They were then washed again in distilled water with removal of the meristematic region and stained with acetic orcein 2%. The analysis of these blades was performed under a 40X magnification optical microscope, taking into account the phase of the cell cycle in which the cells were present, such as interphase, pro-

Table 1. Treatments evaluated in the *Allium cepa* test.

Treatments
T1. Distilled water - Negative control.
T2. Glyphosate 2% - Positive control.
T3. Infusion of 2.5 g of dried leaves.
T4. Infusion of 5.0 g of dried leaves.
T5. Decoction of 2.5 g of dried bark.
T6. Decoction of 5.0 g of dried bark.
T7. Decoction of 2.5 g of dried roots.
T8. Decoction of 5.0 g of dried roots.

phase, metaphase, anaphase and telophase. To calculate the mitotic index was considered the sum of the number of cells in prophase, metaphase, anaphase and telophase, divided by the total number of cells observed (Sehgal et al. 2006; Vieira et al. 2009). The result is presented as a percentage.

2.5. Statistical analysis

The data related to the mitotic index were submitted to the Chi-square test (χ^2) with the aid of the statistical program BioEstat 5.3 (Ayres et al. 2007).

3. RESULTS

The results found for the controls and treatments evaluated in relation to the mitotic index (MI) are presented in Table 2. The cells observed for the counting and evaluation of the treatments were in interphase and cell division (Figure 1). The negative control (T1) presented higher MI (6.12%), differing significantly from all other treatments evaluated. The positive control (T2) also differed statistically from the negative control, showing lower MI, thus confirming its antiproliferative action. Results differed significantly between treatments and controls (Table 2). In the comparison between the negative control (T1) (MI= 6.12%) and the positive control (T2) (MI= 4.55%), there was a significant difference and decreased IM, which indicates inhibition of cell division.

Positive control (T2) differed significantly from treatments with bark extract (T5 and T6) and root

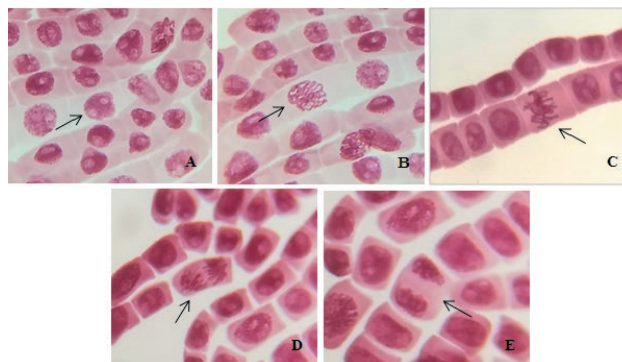


Figure 1. Cell cycle phases of *Allium cepa* with interphase cell and division cells. Interphase (A). Prophase (B). Metaphase (C). Anaphase (D). Telophase (E). Scale 10 μ m.

extract (T7 and T8), and the extracts further inhibited cell division in relation to glyphosate. Comparing the treatments of aqueous extracts of leaves at both concentrations (T3 and T4) with glyphosate treatment (T2), there was the same behavior, as the MI did not differ statistically, showing a decrease in cell division. Treatments with aqueous extracts of dried bark were significantly different. The extract with higher concentration (T6) strongly inhibited cell division, being the lowest mitotic index observed (2.8%). In relation to treatments with dried root extracts, the two differed statistically from each other, but unlike what occurred with treatments with bark extracts, it was the treatment with the lowest concentration (T7) that most reduced the MI (2.95 %).

4. DISCUSSION

All treatments differed significantly from the negative water control (T1) demonstrated by the decrease in MI values. This means that there has been a reduction in cell division of the meristematic cells of *Allium cepa*. This decrease indicates antiproliferative activity of aqueous extracts from the leaves, bark and roots of cabreúva at both concentrations used. The observed cells of the onion bulb root that were submitted to the lowest concentrations of extracts prepared by infusion (T3) and decoction (T5) obtained a cellular stimulus, resulting in the increase of the IM compared to those with higher concentrations (T4 and T6). This means that the higher concentrations of leaf and bark extracts caused the reduction of MI. In contrast, the aqueous extract from dried roots, at higher concentration (T8), increased the MI compared to the lower concentration treatment (T7). Therefore, aqueous extracts of dried leaves and bark

Table 2. Interphase, cell division and mitotic index in *Allium cepa* cells.

Treatments	Cells analyzed	Interphase cells	Dividing cells	MI (%)
T1. Distilled water	4000	3.755	245	6.12 a*
T2. Glyphosate 2%	4000	3.818	182	4.55 b
T3. 2.5 g de DL	4000	3.811	189	4.72 b
T4. 5.0 g de DL	4000	3.816	184	4.60 b
T5. 2.5 g de DB	4000	3.853	147	3.67 c
T6. 5.0 g de DB	4000	3.888	112	2.80 d
T7. 2.5 g de DR	4000	3.882	118	2.95 d
T8. 5.0 g de DR	4000	3.858	142	3.55 c
Total	32000	30681	1319	-

*Averages followed by the same letter in the column do not differ from each other by the χ^2 test at a 5% error probability level. DL= dried leaf; DB= dried bark; DR= dried root; MI= mitotic index.

show antiproliferative capacity, especially at high concentrations.

By studying *Luehea divaricata* in two populations and at concentrations of 6 g L⁻¹ and 30 g L⁻¹, Frescura et al. (2012) also observed this same behavior regarding cell proliferation. The higher concentration led to a decrease in MI values, concluding that there was an increase in antiproliferative capacity with increasing concentration. Coelho's research (2013), analyzing two populations of *Echinodorus grandiflorus* at concentrations of 6 g L⁻¹ and 24 g L⁻¹, also found increased antiproliferative activity at the highest concentration, except for a commercial extract treatment. In this study, regarding the aqueous extracts of leaves, there was no significant difference between the concentrations studied (Table 2). Similarly, Kuhn (2015), when analyzing the *Peltodon longipes* species, observed that leaf extracts at different concentrations (5 g L⁻¹ and 15 g L⁻¹) did not differ significantly. Other tree species such as aroeira (*Myracrodruon urundeuva*) (Trentin et al. 2013), graviola (*Annona muricata*), ipê-roxo (*Handroanthus impetiginosus*) (Melo et al. 2010), angico-branco (*Anadenanthera colubrina*) (Lima et al. 2014) and pau-ferro (*Libidibia ferrea*) (Guerra et al. 2017) also have antiproliferative activity found in their extracts from various plant parts, such as bark and leaves. Some species even belong to the same family as *Myrocarpus*, reaffirming the results obtained in this study.

Myrocarpus frondosus bark extracts had an MI of 3.67% at a concentration of 2.5 g, while the highest concentration had an MI of 2.8%, reducing cell proliferation when the concentration of the extract increased. There was a significant difference between the two concentrations (Table 2). However, other authors, such as Frescura et al. (2012), studying the species *Luehea divaricata*, did not observe significant differences between bark extracts at concentrations 32 g L⁻¹ and 160 g L⁻¹. Considering the results observed for the root decoctions at different concentrations, there is a difference between the concentrations, because the inhibition of cell division was smaller as the extract concentration increased (Table 2). Studying aqueous extracts of *Lavandula angustifolia* roots at a concentration of 0.29 g in 250 mL of distilled water, Freitas et al. (2016) found antiproliferative potential compared to controls. However, Rodrigues et al. (2017), analyzing the same species in higher concentration (3.75 grams in 200 mL of distilled water) observed proliferative potential. The increased concentrations of aqueous extracts of *Lavandula angustifolia* roots induced an increase in the proliferative capacity of the species, a behavior also observed in this study with *Myrocarpus frondosus* roots.

When studying the roots of *Myrocarpus frondosus* *in vivo* and *in vitro*, Bottamedi et al. (2018) found antioxidant and anti-inflammatory activity which were attributed to the presence of flavonoids and phenolics. In addition, the analyzes performed on the essential oil of *Myrocarpus frondosus* leaves showed the presence of α -thujene, α -pinene, sabinene, β -pinene, mircene, p-cymene, limonene, β -bourbonene, β -caryophyllene, D-germacrene, bicyclogermacrene, spatulenol and globulolem, with predominantly β -pinene and bicyclogermacrene (Cabrera et al. 2012; Santi et al. 2017). In other studies with *Echinodorus grandiflorus* (Coelho, 2013), *Baccharis trimera*, *Baccharis articulata* (Fachinetto and Tedesco, 2009), *Caesalpinia echinata* (Bastos et al. 2011) and *Myracrodruon urundeuva* (Romano et al. 2013) were also found substances like flavonoids and tannins. These chemical components were attributed to the antiproliferative capacity presented by the researched plants mentioned above. Flavonoids exert a broad spectrum of health-beneficial biological activities, including the antiproliferative effect on cancer cells (Gibellini et al. 2011; Tsai et al. 2016), reaffirming the results obtained with the species *Myrocarpus frondosus*.

5. CONCLUSION

The aqueous extracts of leaves, bark and roots of *Myrocarpus frondosus* at concentrations of 2.5 and 5.0 grams have antiproliferative effect on the cell division of *Allium cepa* meristematic cells.

Aqueous extracts from the 5.0 gram *Myrocarpus frondosus* bark exhibit high antiproliferative capacity.

The antiproliferative activity found in the species *Myrocarpus frondosus* may be associated with the presence of flavonoid and phenolic compounds in the plant tissue composition of the species.

ACKNOWLEDGMENTS

To CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico) and Capes (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) for the funding of the research project.

REFERENCES

- Ayres M, Ayres Júnior M, Ayres DL, Santos AS. 2007. Bioestat 5.0: aplicações estatísticas nas áreas das ciências biológicas e médicas. Belém, PA, Brasil.

- Bastos IVGA, Silva GKC, Rodrigues GCR, Melo CM, Xavier HS, Souza IA. 2011. Estudo fitoquímico preliminar e avaliação da toxicidade aguda do extrato etanólico bruto de *Caesalpinia echinata* Lam. Rev. Bras. Farm. 92: 219-222. <http://www.rbfarma.org.br/files/rbf-2011-92-3-23.pdf>
- Bottamedi M, Nascimento MVPS, Fratoni E, Moon YJK, Dalmarco EM, Mendes BG. Evaluation of antioxidant and anti-inflammatory action (*in vivo* and *in vitro*) from the trunk barks of *Myrocarpus frondosus* Allemão (Cabreúva). In: 25° Simpósio de Plantas Mediciniais do Brasil, São Paulo/Brasil, 2018. http://www.eventus.com.br/plantasmediciniais2018/anais_xxv_simposio_plantas_mediciniais_2018.pdf
- Cabrera DC, Gomes GLS, Schmidt N, Flach A, Costa LAMA, Rosa GR, Moura NF. 2012. Composição química das folhas da espécie *Myrocarpus frondosus* do sul do Brasil. In: 35ª Reunião Anual da Sociedade Brasileira de Química, São Paulo/Brasil. <http://sec.sbq.org.br/cdrom/35ra/resumos/T0374-1.pdf>
- Caritá R, Marin-Morales MA. 2008. Induction of chromosome aberrations in the *Allium cepa* test system caused by the exposure of seeds to industrial effluents contaminated with azo dyes. Chemosphere. 72: 722-725. <https://doi.org/10.1016/j.chemosphere.2008.03.056>
- Coelho APD. 2013. Potencial genotóxico e antiproliferativo dos extratos de *Echinodorus grandiflorus* e *Sagittaria montevidensis* (Alismataceae). [Dissertação] - Universidade Federal de Santa Maria, Santa Maria, RS, Brasil.
- Fachinetto JM, Tedesco SB. 2009. Atividade antiproliferativa e mutagênica dos extratos aquosos de *Baccharis trimera* (Less.) e *Baccharis articulata* (Lam.) Pers. (Asteraceae) sobre o sistema teste de *Allium cepa*. Rev. Bras. Plantas Med. 11: 360-367. <https://doi.org/10.1590/s1516-05722009000400002>
- Freitas JMB, Tedesco SB, Rodrigues LG, Pasqualli M. 2016. Avaliação do potencial antiproliferativo do extrato de *Lavandula angustifolia* pelo teste de *Allium cepa*. In: 31° Jornada Acadêmica Integrada, Santa Maria/RS/Brasil, 2016. <https://portal.ufsm.br/jai/trabalho/trabalho.html?action=anais>
- Frescura VD, Laughinghouse IV HD, Tedesco SB. 2012. Antiproliferative effect of the tree and medicinal species *Luehea divaricata* on the *Allium cepa* cell cycle. Caryologia. 65(1): 27-33. <https://doi.org/10.1080/00087114.2012.678083>
- Gibellini L, Pinti M, Nasi M, Montagna JP, Biasi S, Roat E, Bertoncelli L, Cossarizza A. 2011. Quercetin and cancer chemoprevention. Evid.-Based Complementary Altern. Med. 1-15. <https://doi.org/10.1093/ecam/neq053>
- Guerra ACVA, Soares LAL, Ferreira MRA, Araújo AA, Rocha HAO, Medeiros JS, Cavalcante RDS, Araújo Júnior RF. 2017. *Libidibia ferrea* presents antiproliferative, apoptotic and antioxidant effects in a colorectal cancer cell line. Biomed. Pharmacother. 92: 696-706. <https://doi.org/10.1016/j.biopha.2017.05.123>
- Guerra M, Souza MJ. 2002. Como observar cromossomos: um guia de técnicas em citogenética vegetal, animal e humana. Fundação de Pesquisas Científicas de Ribeirão Preto, Ribeirão Preto, SP, Brasil.
- Kuhn AW. 2015. Viabilidade polínica, genotoxicidade, efeito antiproliferativo e compostos fenólicos de *Peltodon longipes* Kunth Ex Benth. (Lamiaceae). [Dissertação] - Universidade Federal de Santa Maria, Santa Maria, RS, Brasil.
- Leme DM, Marin-Morales MA. 2009. *Allium cepa* test in environmental monitoring: A review on its application. Mutat. Res. 682(1): 71-81. <https://doi.org/10.1016/j.mrrev.2009.06.002>
- Lima RDE, Alves É P, Rosalen PL, Ruiz ALTG, Duarte MCT, Góes VFF, Medeiros ACD, Pereira JV, Godoy GP, Costa EMMB. 2014. Antimicrobial and antiproliferative potential of *Anadenanthera colubrina* (Vell.) Brenan. Evid.-Based Complementary Altern. Med. 1-7. <https://doi.org/10.1155/2014/802696>
- Lorenzi H, Matos FJA. 2008. Plantas medicinais no Brasil: Nativas e Exóticas. 2ª ed. Nova Odessa - SP, Instituto Plantarum, Brasil.
- Melo JG, Araújo TAS, Almeida Castro VTN, Cabral DLV, Rodrigues MD, Nascimento SC, Amorim ELC, Albuquerque UP. 2010. Antiproliferative activity, antioxidant capacity and tannin content in plants of semi-arid northeastern Brazil. Molecules. 15(12): 8534-8542. <https://doi.org/10.3390/molecules15128534>
- Pereira Júnior LR, Andrade AP, Araújo KD, Barbosa AS, Barbosa FM. 2014. Espécies da Caatinga como alternativa para o desenvolvimento de novos fitofármacos. Floram. 21: 509-520. <https://doi.org/10.1590/2179-8087.024212>
- Rodrigues LG, Tedesco SB, Trapp KC, Rosa VS. 2017. Efeito proliferativo de extratos aquosos de *Lavandula angustifolia* sob o ciclo celular de *Allium cepa*. In: 32° Jornada Acadêmica Integrada, Santa Maria, RS, Brasil. <https://portal.ufsm.br/jai/trabalho/trabalho.html?action=anais>
- Romano B, Pagano E, Montanaro V, Fortunato AL, Milic N., Borrelli F. 2013. Novel insights into the pharmacology of flavonoids. Phytoter. Res. 27(11): 1588-1596. <https://doi.org/10.1002/ptr.5023>
- Santi II, Gatto DA, Machado MRG, Santos PSB, Freitag RA. 2017. Chemical composition, antioxidant and antimicrobial activity of the oil and plant extract

- Myrocarpus frondosus* Allemão. Am. J. Plant Sci. 8(7): 1560-1571. <https://doi.org/10.4236/ajps.2017.87108>
- Scoti MSV, Araujo MM, Wendler CF, Longhi SJ. 2011. Mecanismos de regeneração natural em remanescente de Floresta Estacional Decidual. Ci. Fl. 21: 459-472. <https://doi.org/10.5902/198050983803>
- Sehgal R, Roy S, Kumar VL. 2006. Evaluation of cytotoxic potential of latex of *Calotropis procera* and *Podophyllotoxin* in *Allium cepa* root model. Biocell.30(1): 9-13. PMID: 16845823
- Tedesco SB, Laughinghouse IV HD. 2012. Bioindicator of genotoxicity: the *Allium cepa* test. In: Environmental Contamination, InTech Publisher. 137-156. <https://doi.org/10.5772/31371>
- Trentin DS, Silva DB, Amaral MW, Zimmer KR, Silva MV, Lopes NP, Giordani RB, Macedo AJ. 2013. Tannins possessing bacteriostatic effect impair *Pseudomonas aeruginosa* adhesion and biofilm formation. Plos One. 8(6): 1-13. <https://doi.org/10.1371/journal.pone.0066257>
- Tsai PH, Cheng CH, Lin CY, Huang YT, Lee LT, Kandaswami CC, Lin YC, Lee KP, Hung CC, Hwang JJ, Ke FC, Chang GD, Lee MT. 2016. Dietary flavonoids luteolin and quercetin suppressed cancer stem cell properties and metastatic potential of isolated prostate cancer cells. Anticancer Res. 36(12): 6367-6380. <https://doi.org/10.21873/anticancer.11234>
- Ubessi C, Tedesco SB, Silva CB, Baldoni M, Kryszczun DK, Heinzmann BM, Rosa IA, Mori NC. 2019. Antiproliferative potential and phenolic compounds of infusions and essential oil of chamomile cultivated with homeopathy. J. Ethnopharmacol. 239: 1-7. <https://doi.org/10.1016/j.jep.2019.111907>
- Vieira A, Guimarães MA, David GQ, Karsburg IV, Campos ANR. 2009. Efeito genotóxico da infusão de capítulos florais de camomila. R. Trop.: Ci. Agr. Biol. 3: 8-13. https://www.researchgate.net/publication/246044615_Evaluation_of_genotoxic_effects_of_chamomile_floral_chapters_infusion_Efeito_genotoxico_da_infusao_de_capitulos_florais_de_camomila