



**Citation:** F. Fallah, F. Ghahremaninejad (2021) Ploidy level determination of *Hedera* (Araliaceae) with an emphasis on discussable species (*Hedera hibernica*). *Caryologia* 74(1): 109-116. doi: 10.36253/caryologia-881

**Received:** March 19, 2020

**Accepted:** October 02, 2020

**Published:** July 20, 2021

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**Data Availability Statement:** All relevant data are within the paper and its Supporting Information files.

**Competing Interests:** The Author(s) declare(s) no conflict of interest.

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## Ploidy level determination of *Hedera* (Araliaceae) with an emphasis on discussable species (*Hedera hibernica*)

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**Abstract.** Genome size is a helpful tool for circumscribing taxa at diverse taxonomic degrees (mostly species) and resolving intricate low-level taxonomies. The correct genome size in *Hedera* (Araliaceae) has long been discussed, and the ploidy levels of some taxa are still unclear. Twelve accessions of *Hedera* were measured via flow cytometry. Flow cytometry is a relatively rapid, inexpensive, and credible tool. Fresh leaves of *Hedera* samples and internal reference standard parsley (*Petroselinum crispum*) were stained with propidium iodide (PI). Flow cytometry measurements showed that for the accessions of 2CV (3.09 - 6.40 pg), the lowest amount of nuclear DNA was 3.09 pg for *Hedera crebescens* (So), while the highest amount was 6.40 pg for *H. hibernica* "Hamilton," representing a statistically significant difference. According to this study, the new taxon (*H. crebescens*) is a diploid, though this taxon was previously considered *H. hibernica* (tetraploid).

**Keywords:** flow cytometry, *Petroselinum crispum*, genome size, *Hedera crebescens*.

### INTRODUCTION

*Hedera* L. is an evergreen woody vine native to Europe, Asia, and North Africa, but it is cultivated worldwide (Rose 1996; Reichard 2000; Ackerfield and Wen 2002, 2003). The taxonomic history of the *Hedera* taxa is complicated because of its two-part life cycle, extensive geographic distribution. Juvenile phase with palmately lobed leaves on sterile stems and adult phase with unlobed cordate adult leaves on fertile flowering stems. The juvenile and adult shoots also differ, the juvenile being slender, pliable and scrambling or climbing with small aerial roots to fix the shoot to the substrate (rock or tree bark), the adult shoots thicker and without aerial roots. (Rose 1996; Rutherford et al. 1993; Ackerfield and Wen 2002).

Taxonomic recognition was first afforded to juvenile and adult plants, which were described by Linnaeus (1753) as "*H. helix* L." and "*H. arborea* (L.) Walter." A previous investigation on *Hedera* recognized five species (Lawrence and Schulze 1942). However, later studies on *Hedera* have suggested that these five species should be subdivided into more species (Ackerfield 2001).

Furthermore, a key to the classification of species and subspecies of *Hedera* has been derived based on trichome and leaf morphology (Ackerfield and Wen 2002). Recently the European Garden Flora has reported that there are 12 *Hedera* taxa (McAllister and Marshall 2017). However, the definition of species and identification of taxa are still disputed (Valcárcel and Vargas 2010).

Since ancient times, many *Hedera* cultivars have been used in Europe for cover green and garden decorations (Rose 1996) and a remarkable number of cultivars have been identified as *H. hibernica* (G. Kirchn.) Bean and *H. helix* subsp *hibernica* (G. Kirchn.) (D.C. McClinton). Based on differences in trichome positioning, leaf form, and chromosome number, *H. hibernica* has been recognized (McAllister and Rutherford 1990) as a distinct species from *H. helix*. It has been reported (McAllister and Rutherford 1990; Jacobsen 1954) that *H. helix* and *H. hibernica* have different chromosome numbers, with *H. helix* being diploid ( $2n=48$ ) and *H. hibernica* being tetraploid ( $2n=96$ ).

*H. hibernica* was thought to be a unique tetraploid species, and hence, it was carefully compared with a typical diploid *H. helix*. However, recent molecular data indicate that *H. helix* and *H. hibernica* may represent different species (Metcalf 2005). *H. helix* (diploid) has been the maternal parent for the tetraploid *H. hibernica* (Ackerfield and Wen 2003). Sometimes, diploid and polyploid species of the *Hedera* genus are barely distinct morphologically or through DNA sequencing data (Green et al. 2011). The taxonomic and evolutionary significance of variations in genome size has been established, and chromosomal data are extensively used in plant taxonomy (Stace 2000; Kron et al. 2007; Ekrt et al. 2009). Increasing ploidy usually results in increased cell size. Plants with increased ploidy levels may be apparently distinct morphologically.

Flow cytometry is a very useful tool for measuring DNA content and can be related to the ploidy level for a specified taxon (Sharma and Sharma 1999). Although genome size in *Hedera* has long been disputed, the genomic DNA amounts of many taxa are still unknown (Domoney and Timmis 1980; Polito and Alliata 1981; König et al. 1987).

However, molecular studies based on flow cytometric measurements have revealed that polyploidy has been significant to the evolution of the *Hedera* species and might have taken place many times independently in various lineages (Green et al. 2011).

Recent reports have indicated that some taxa with different morphological and cytological characteristics are spreading in semi-natural habitats and urban areas that contain escaped gardens (Udvardy and Bényei-Himmer 1999).

Recently, by studying ivy diversity in Hungary, researchers have identified a prominent *Hedera* taxon that has a particular habit, contains a set of distinguishable morphological and phonological features, and has various environmental demands. Previously, this was thought to be *H. hibernica* (Bényei-Himmer et al. 2017). However, Bényei and Höhn (2017) recently identified this taxon as *H. crebescens*. The purpose of the present study was to use flow cytometry measurements to clarify the situation of the *Hedera* taxa that is spreading in Hungary and other countries in central Europe, was previously identified as *H. hibernica*, and has been recently identified as *H. crebescens*.

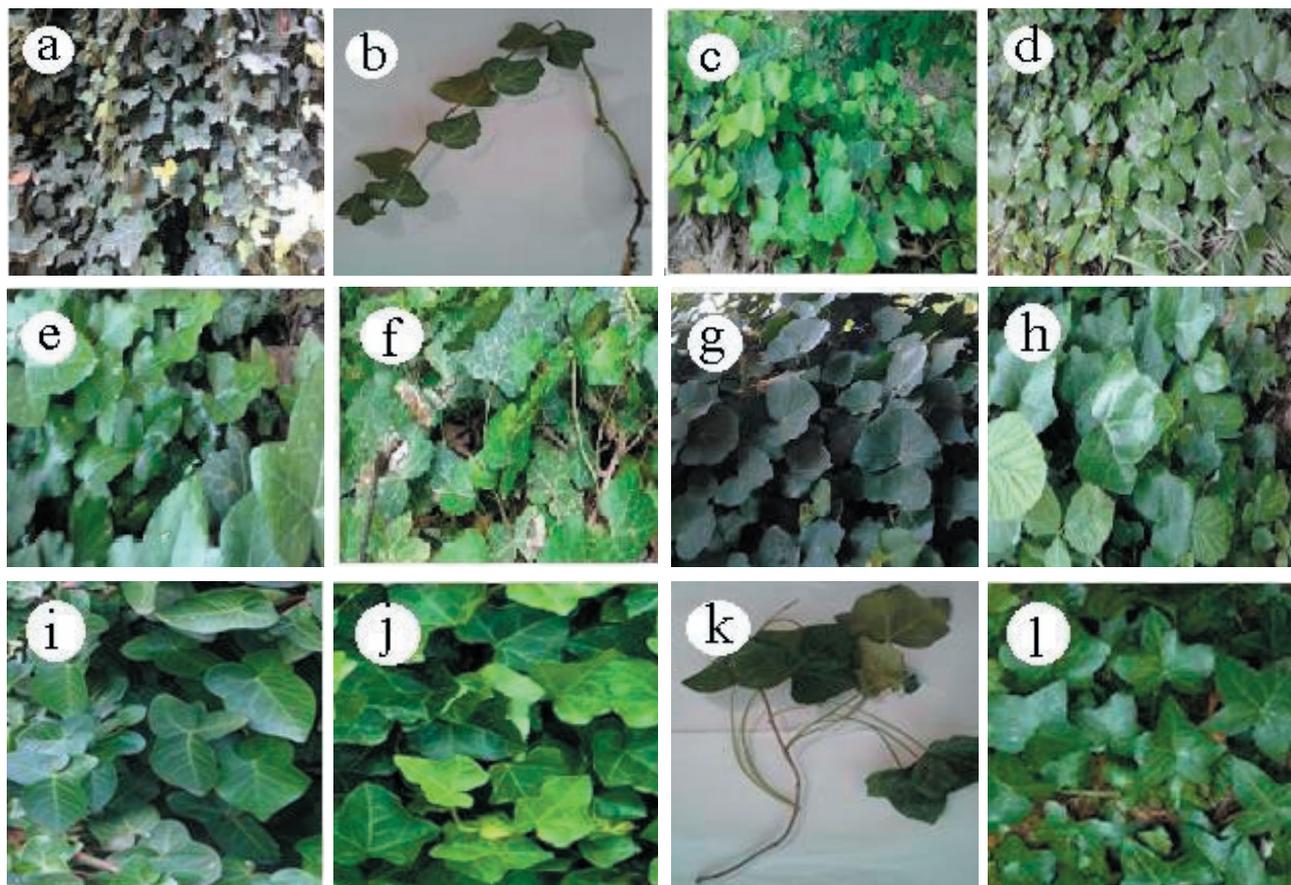
## MATERIALS AND METHODS

*Hedera* specimens were collected from different natural habitats in Central Europe and from the Soroksár Botanical Garden of Budapest (Fig. 1). Genome size was examined by flow cytometry. Following Ramsey (2007) completely expanded fresh leaf tissues from each sample (0.5 g) were manually chopped with a sharp razor blade for approximately 1 min in petri dish with the same amount of leaf tissue from diploid *Petroselinum crispum* (Apiaceae) as an internal reference standard (Fig. 2), in 2 mL of isolation buffer (3.6 g HEPES, 2 mL of a 0.5 M solution of EDTA, 6.0 g KCL, 1.2 g Na Cl, 102.7 g sucrose, 2 mL Triton X-100, 1 mL  $\beta$ -mercaptoethanol, and 0.1 g spermine in 1.0 L distilled water). Following that, nuclear suspension was filtered through a nylon mesh (25  $\mu$ m) to remove debris and then stained with Propidium Iodide (PI) at a final density 100  $\mu$ g. mL<sup>-1</sup> and complemented with 100  $\mu$ g. mL<sup>-1</sup> ribonuclease A (RNAs). IC-value was calculated based on the converting formula (Dolezel et al. 2003) [ $1pg=978$  mega base pairs (Mbp)].

The relationship between mean ( $n=3$ ) 2C-values of leaf samples were processed and the resulting fluorescence histograms were analyzed with Flomax Software. The total DNA amount of a sample was calculated based on the values of the G1 peak means as follows (Dolezel et al. 2003, 2007; Dolezel and Bartos 2005.) (Sample 2C DNA (pg) content= [(Sample G1 peak mean) / (Standard G1 peak mean)] $\times$ 2C DNA amount Standard).

## RESULTS AND DISCUSSION

As presented in Table 1, 12 taxa of *Hedera* genus were evaluated by Partech Flomax software Ver. 2.0.01 in order to assess the nuclear DNA contents (pg) and genome sizes (Mbp). Among 8 diploids examined, the



**Figure 1.** Images of *Hedera* species analysed in this study: a,b: *H. helix*, c: *H. helix arborescence*, d,e: *H. hibernica*, f: *H. hibernica arborescence*, g,h: *H. crebescens*, i: *H. hibernica* “Deltoideadea”, j, k: *H. hibernica* “Variegata”, l: *H. hibernica* “Hamilton”.

lowest amount of nuclear DNA was 3.09 pg for *H. crebescens* (So) and 3.33 pg for the *H. crebescens* (JH). As a result, a statistically insignificant difference of 0.24 pg was observed between four tetraploids while a statistically significant difference in 2C-value (0.64 pg) in the range of 5.76-6.40 pg is noticed between *H. hibernica* “Variegata” (5.76 pg) and *H. hibernica* “Hamilton” (6.40 pg).

Determination of genome size and ploidy level are summarized in (Table 1) and results analyzing the amount of nuclear DNA are shown in (Fig. 3, 4). Sometimes in *Hedera* species the related diploid and polyploidy are poorly distinct via morphology and DNA sequence data (Green et al. 2011).

Although polyploidy in the *Hedera* genus is common, the occurrence of auto and allopolyploidy is poorly understood (Yi et al. 2004). Trichome morphology and leaf shape analyses revealed that *H. hibernica* is an allopolyploid from *H. helix* and *H. maroccana* (McAllister and Rutherford 1990). Furthermore, based on nucleotide polymorphisms in nrDNA (ITS) *H. hibernica* was

recognized as an allopolyploid (Vargas et al. 1999). It has commonly been believed that internal reduplication is correlated with very small genomes, assuming that minimal DNA content is needed for suitable cell functioning. In fact, taxonomists have surely realized that some related species with the same number of chromosomes might be different in terms of DNA volume, thus making them easily distinctive by using flow cytometry.

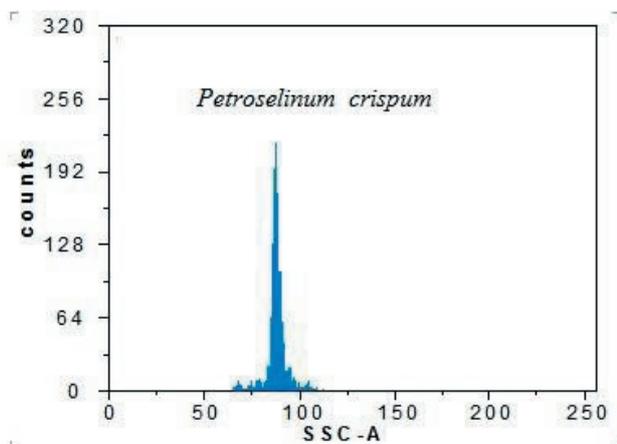
Recent studies on vascular plants revealed only a weak negative correlation between genome size and degree of polyploidy (Barow and Meister 2005). Thus, C-values should be treated as a fundamental scaling factor in living systems (Bennett et al. 2000).

The specimens examined in the present work by flow cytometry measurements showed that the accessions 2C-value (3.06-6.40 pg) verifying more than a two-fold variation and showing a corresponding genome size of 1516-3129 Mbp. Among the eight examined diploids, the lowest amount of nuclear DNA was 3.09 pg (for *H. crebescens* (So)), while the highest amount was

**Table 1.** Samples, locality and determination of genome size and ploidy level.

Samples	Locality	Latitude ( N )	Longitude ( E )	Ploidy level	Mean 2C value level (pg± SE)	Mean 1C value (pg)	Mean 1C value (Mbp)*	% CV
<i>H. helix</i>	Soroksár (Hungary)	N 47°23' 18.644"	E 19°9' 2.165"	2x	3.16 ± 0.05	1.58	1545.24	0.02
<i>H. helix arborescence</i>	Gellért Hill (Hungary)	N 47°29' 12.898"	E 19°2' 40.23"	2x	3.16± 0.04	1.58	1545.24	0.01
<i>H. hibernica</i>	Gellért Hill (Hungary)	N 47°29' 12.898"	E 19°2' 40.23"	4x	6.23 ± 0.30	3.11	3041.58	0.05
<i>H. hibernica arborescence</i>	Soroksár (Hungary)	N 47°23' 18.644"	E 19°9' 2.165"	2x	3.17 ± 0.07	1.57	1535.46	0.02
<i>H. hibernica</i> "Deltoiedea"	Soroksár (Hungary)	N 47°23' 18.644"	E 19°9' 2.165"	4x	6.33± 0.12	3.16	3090.48	0.02
<i>H. hibernica</i> "Hamilton"	Gellért Hill (Hungary)	N 47°29' 12.898"	E 19°2' 40.23"	4x	6.40± 0.07	3.2	3129.6	0.01
<i>H. hibernica</i> "Variegata"	Soroksár (Hungary)	N 47°23' 18.644"	E 19°9' 2.165"	4x	5.76± 0.11	2.88	2816.64	0.02
<i>H. crebescens</i> (V)	Vienna (Austria)	N 48°12' 31.307"	E 16°22' 21.702"	2x	3.19 ± 0.01	1.59	1555.02	0.01
<i>H. crebescens</i> (JH)	János Hill (Hungary)	N 47°29' 52.836"	E 19°2' 23.675"	2x	3.33 ± 0.12	1.66	1623.48	0.04
<i>H. crebescens</i> (Sze)	Szeged (Hungary)	N 46°10' 18.332"	E 19°25' 22.52"	2x	3.22 ± 0.05	1.61	1574.58	0.02
<i>H. crebescens</i> (So)	Soroksár (Hungary)	N 47°23' 18.644"	E 19°9' 2.165"	2x	3.09± 0.07	1.54	1506.12	0.02

\*1 pg = 978 Mbp<sup>27</sup>



**Figure 2.** The histogram for analysis of the amount of nuclear DNA in leaves: parsley (*P. crispum*) reference standard (2C DNA=4.50 pg).

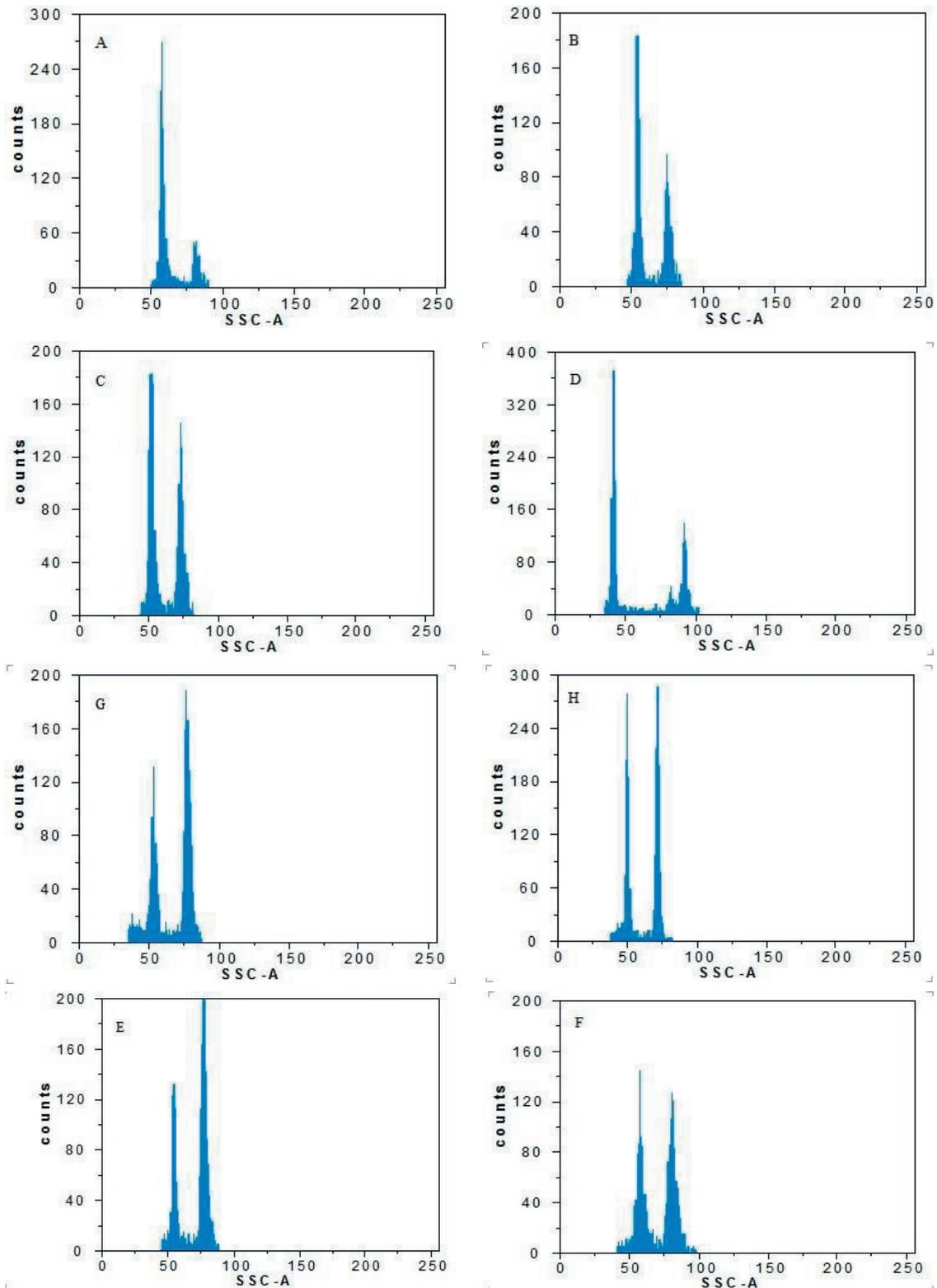
3.33 pg (for *H. crebescens* (JH)). Statistically in significant differences were found between four tetraploids. Meanwhile, significant differences of 0.64 pg in 2C-value (ranging from 5.76-6.40 pg) were recognized for *H. hibernica* "Variegata" (5.71 pg) and *H. hibernica* "Hamilton" (6.40 pg).

The results attained by Marie and Brown (1993) indicate that for *H. helix*, 2CV=8.18 pg, which is strongly refuted by the data presented in the current study and stands as an uncommon value for DNA amounts (Bennett and Leitch 1997). *Petunia hybrid* Vilm. was used by Marie and Brown (1993) as a standard reference. *H. helix* which studied by Marie and Brown from Stras-

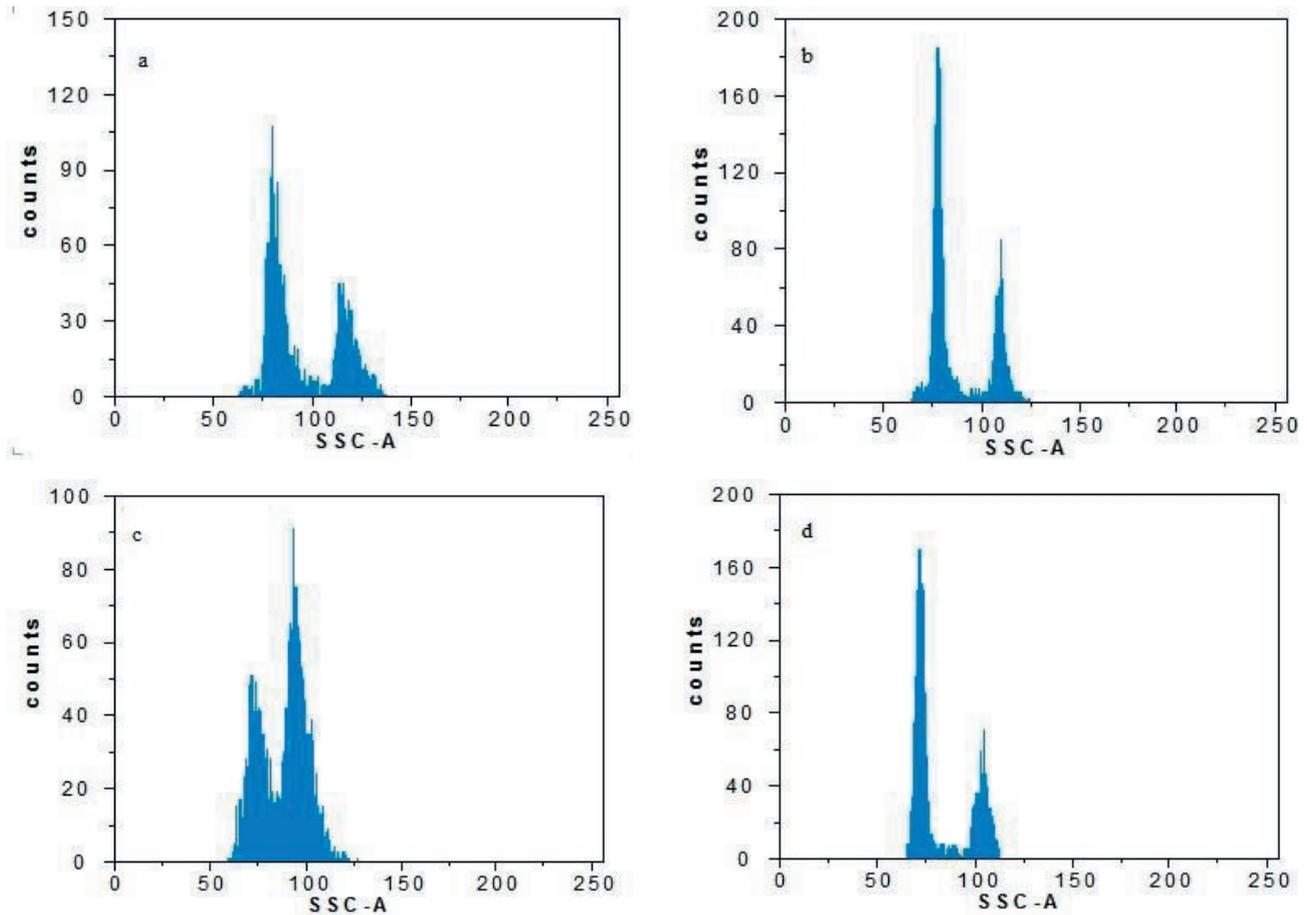
burg-France (Latitude N 48°34'50.959", Longitude E 7°45'49.623") is not very far from to region we were collected *H. helix*. According to the present study, the data for *H. helix* (2CV=4.6 pg) reported by Domoney and Timmis (1980) remains unsupported.

As reported previously, *H. helix* 2CV=2.95 (Konig et al. 1987), *H. helix* 2CV=2.80 (Obermayer and Greilhuber 2000) and *H. hibernica* 2CV= 6.00 (Zonneveld et al. 2005), which are close to the present results. The morphometric analyses of *H. helix*, *H. hibernica* "Hamilton" and the new taxon (*H. crebescens*) were based on vegetative and generative organs and were conducted to distinguish the new taxon from *H. helix* and *H. hibernica* (Clarke et al. 2006). The genome size, which represents an inherent attribute, is a supportive feature for circumscribing taxa of various taxonomic ratings (mainly species) and resolving intricate low-level taxonomies (Loureiro et al. 2006).

The analysis based on flow cytometry indicated that *H. crebescens* can be considered a distinct taxon among the diploid ivies. According to the present study, which is strongly supported by the notion of a newly identified diploid taxon (*Hedera crebescens*) (Bényei and Höhn 2017), it should be emphasized that this taxon which was previously treated as *H. hibernica* is not identical to the tetraploid taxon.



**Figure 3.** The histogram for analysis of the amount of nuclear DNA in leaves: The left peaks refer to the unknown *Hedera* samples and the right peaks to the known parsley (*P. crispum*) reference standard (2C DNA = 4.50 pg). A: *H. crebescens* (V) = 3.19 pg. B: *H. crebescens* (JH) = 3.33 pg. C: *H. crebescens* (Sze) = 3.22 pg. D: *H. crebescens* (So) = 3.09 pg. E: *H. crebescens* (GH) 5 = 3.16 pg. F: *H. helix*=3.16 pg. G: *H. helix* arborescence=3.16 pg, H: *H. hibernica* arborescence=3.17 pg, diploids *Hedera*.



**Figure 4.** The histogram for analysis of the amount of nuclear DNA in leaves: The left peaks refer to the known parsley (*P. crispum*) reference standard and right peaks to the unknown *Hedera* samples. a: *H. hibernica* = 6.23 pg, b: *H. hibernica* "Deltoidea" = 6.33 pg, c: *H. hibernica* "Variegata" = 5.76 pg, d: *H. hibernica* "Hamilton" = 6.40 p.

## CONCLUSIONS

Most of the recently reported new incidences of *H. helix* from the lowland of Hungary (see Bartha and Király 2015) refer to *H. crebrescens*. *H. crebrescens* is the most invasive ivy taxon in Hungary and probably most of the countries in central Europe. According to flow cytometry results, *H. hibernica* arborescence is diploid, whereas *H. hibernica* is tetraploid. Therefore, this name is not correct for this taxon, and due to its morphological features, it is probably a subspecies of *H. crebrescens*. Studies that include species from the eastern part of the distribution range of the *Hedera* genus in Iran and the Caucasus (formerly mentioned by K. Koch and G. Woronow) are necessary in order to conduct a more thorough survey of *Hedera*'s diversity and relationships.

## ACKNOWLEDGEMENTS

We would like to thank Prof. Ghasem Karimzade, the head of the Laboratory Biotechnology, Faculty of Agriculture, Tarbiat Modares University, Tehran, Iran for his technical assistance. We would also like to extend our gratitude to Prof. Jaroslav Doležel, the head of the Laboratory of Molecular Cytogenetic and Cytometry, Institute of Experimental Botany, Sokolovska' 6, 456 Olomouc, Czech Republic, for providing seeds of DNA reference standards and all his papers.

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